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Traditional Uses and Anticancer Potential of the *Combretum* Genus: A Literature Review

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Author's contribution

The sole author designed, analyzed, interpreted and prepared the manuscript.

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Review Article

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ABSTRACT

Background: Cancer incidence is increasing annually in all countries. So, it is nowadays a great burden for the different nations of the world. Research for new therapeutics is becoming an urgent need, particularly for intractable and chemoresistant cancer cases. The solutions can still be found by investigating natural products which are recognized as promising sources of bioactive compounds with a potential for the discovery of new preventive and therapeutic anticancer agents. **Methodology:** The present work used databases such as Pubmed, Science Direct and Google scholar to investigate the ethnobotanical uses of some *Combretum* species in the literature. It also allowed us to summarize some pharmacological studies on *Combretum* species.

Results: This review gathers all available traditional uses and cytotoxicity studies of *Combretum* species in the literature. Special focus is given to pharmacological studies highlighting isolated potential anticancer molecules. These molecules present potent cytotoxic effect on various cancer cell lines and may contribute to improving the health of people suffering from various cancer diseases.

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Conclusion: The *Combretum* species are widely used in folk medicine for the treatment of several pathologies including cancers. This study is of fundamental importance in highlighting *Combretum* species as a potential source for research of new anticancer compounds.

Keywords: Anticancer potential; combretum; ethnobotanical uses; natural product; traditional medicine.

1. INTRODUCTION

The different cancer diseases continue to have a negative impact on the world. Despite advances in the search for better solutions against the scourge, its incidence is tending to increase. Indeed, statistics show that new cases of cancer increased from 12.7 million in 2008 [1] to 14.1 million in 2012 [2], then to 18.1 million in 2018 [3]. Estimates predict 28.4 million new cases of cancer in 2040 [4]. Cancer caused nearly 10 million deaths in 2020, making cancer one of the leading causes of death worldwide [4]. Cancer is also a major concern for Burkina Faso. In the year 2020, GLOBOCAN estimates revealed 12,045 new cases of cancer, including 8,695 deaths. More than 18,884 new cases are expected for the next five years. Within noncommunicable diseases, cancer is the second most causing death in this country after cardiovascular diseases. The most common cancers are breast (16%), liver (10.3%), cervix uteri (9.4%), prostate (8.3%), and bladder (5.5%) [5]. To fight cancer effectively, the Ministry of Health has adopted a strategic plan for 2021 to 2025 in which strong actions against cancer in Burkina Faso have been undertaken. Among other things, there is the strengthening of cancer research, in particular the identification of new antitumor molecules contained in the country's medicinal plants. Burkina Faso has a very rich and varied traditional pharmacopeia [6,7] to which more than 80% of the population has recourse to it for their essential healthcare needs [8].

Medicinal plants have always been a credible alternative for the search for active molecules to counter major public health scourges in the interest of the survival of populations since antiquity [9]. Active molecules of natural origin have long served as a source of therapeutic agents or as a model for the synthesis of derivatives [10]. As part of the fight against cancer, many anti-cancer molecules have been identified from certain medicinal plants. This is the case of vincristine and vinblastine isolated from species of the genus Catharanthus; paclitaxel isolated from species of the genus podophyllotoxin isolated Taxus: from Podophyllum emodi; Colchicine isolated from species of the genus Colchicum and many other compounds that are currently used clinically for the treatment of patients with cancer [11]. Given the emergence of cancer resistance forms to current treatments, it is important to always look anticancer bioactive for new molecules. Ethnobotanical studies show that Combretum species are used in the treatment of several pathologies including cancers and their cytotoxicity effects on various cancer cell models have been pharmacologically demonstrated [12-15]. These diverse therapeutic properties of Combretum species are due to the wide variety of secondary metabolites contained in these plants. Indeed, phytochemical analyzes have revealed the presence of many classes of constituents, including triterpenes, flavonoids, lignans, alkaloids, and non-protein amino acids, among others in most species of the Combretum genus [16-18]. So, species of the Combretum genus which are widely used in traditional medicine can be an important source of new anticancer molecules discovery.

The objective of this review was to gather all available traditional uses and cytotoxicity studies of *Combretum* species in the literature. Pharmacological studies highlighting isolated potential anticancer molecules from *Combretum* species were reported as well. This study will contribute to highlight *Combretum* species as a potential source for research of new anticancer compounds.

2. METHODOLOGY

PubMed, Science Direct and Google Scholar databases were used to select the articles. For the search in these databases, the following keywords were used: "*Combretum*," "Traditional uses," "Cytotoxicity" and "Cancer". The articles were selected according to these inclusion criteria: made with *Combretum* species; title and summary with traditional uses of combretum species; title and summary with the assessment of cytotoxicity activity of *Combretum* extracts on cancer cell lines; title and summary with the evaluation of cytotoxicity activity of isolated compound from *Combretum* extracts. Exclusion criteria were: duplication of articles and abstracts and full texts irrelevant to the topics in question. Some books and Ph.D. thesis on the topics were also consulted.

3. RESULTS AND DISCUSSION

3.1 Ethnobotanical Uses of *Combretum* Species

Traditional medicine has a prominent place in the world and specifically in Sub-Sahara developing countries [19]. Indeed, with the resurgence of diseases non-accessibility and the of pharmaceutical drugs, populations living in lowincome regions of the world are looking to folk medicine for their healthcare needs [20]. Medicinal plants are mostly used by traditional healers for therapeutic purposes [21]. Ethnobotanical studies have been very important hiahliahtina some traditional in medical knowledge of indigenous peoples and traditional healers. which has contributed to the establishment of a list of medicinal plants used in folk medicine as the first line of primary healthcare in the rural community. Such information is useful for researchers looking for new bioactive compounds, especially anti-cancer agents.

The Combretum genus belonging to the Combretaceae family includes nearly 250 plant species among which 12 species are represented in Burkina Faso [22]. Species of Combretum are well spread across Sub-Sahara Africa. The geographical distribution of Combretum species in Burkina Faso has been investigated, and results showed that these species are widespread on the territory [23]. A positive correlation has been observed by some authors between the accessibility of plants and their medicinal uses [24]. Thus, the wide distribution of Combretum species across Africa and particularly on the territory of Burkina Faso could explain the medicinal knowledge of these plants and consequently their wide use by populations.

Combretum species are variously used for the treatment of several pathologies. An ethnobotanical study revealed that different parts of C. molle are used by traditional healers in Uganda for the treatment of various cancers such as abdominal, bone, cervical, intestinal, liver, skin, throat cancers, and leukemia Another ethnobotanical [25]. investigation realized in Nigeria points out three Combretum species such as C. micranthum, C. Camporum, and C. molle that are used by

traditional healers for the treatment of cancers [14]. The roots and leaves of *C. zeyheri* are used by traditional healers in Tanzania for the treatment of cancer [12]. A study reported the traditional uses of *C. fragrans* in the northern part of Cameroon for the treatment of cancers [15]. Others traditional uses of some *Combretum* species recorded in the literature are showed in Table 1.

Medicinal plants from Burkina Faso are also well used by populations for their healthcare needs. Some ethnobotanical studies allowed to highlight the traditional knowledge of traditional healers in the country. Indeed, some researchers have listed 61 species of medicinal plants used in the care of kidney diseases in Burkina Faso, among which C. micranthum was mentioned [26]. Another study reported nearly 134 medicinal recipes from 106 species of medicinal plants from Burkina Faso including C. molle for their various medicinal uses [27]. Also. an showed that 94 species investigation of medicinal plants are used to combat different pathologies in Burkina Faso of which C. glutinosum, C. micranthum, C. nigricans, and C. paniculatum are cited [28]. To the best of our study highlighting knowledge, а the ethnobotanical uses of Combretum species for cancer treatment in Burkina Faso is not yet reported.

These *Combretum* species could be an important resource for researchers, especially to those from Burkina Faso where the cancer healthcare system and chemotherapy are not accessible, to investigate their anticancer potential and to isolate new anticancer molecules as well.

3.2 Anticancer Potential of *Combretum* Species

Cancer is a great burden for the different nations of the world. This has led different nations to adopt strategic plans to overcome this pathology. Also, the fact that resistance to available treatments is increasingly appearing motivate researchers to look for new therapeutic molecules and even new therapeutic targets for therapy. Species of the cancer genus *Combretum* could play a key role in the discovery of new therapeutic molecules against cancer. Indeed, some species of this genus are well known for their traditional uses against cancer. Also, several pharmacological studies have been undertaken to elucidate the anticancer properties of plant species from the Combretum genus.

Species	Used parts	Traditional uses	References
<i>C. aculeatum</i> Vent.	Leafy stems, roots	Diarrhea, gonorrhea, intestinal parasites, colic, dysentery, jaundice, malaria, hypertension, fevers, constipation, fractures, weight delays, hypocalcemia, remineralizing, spasmodic, mental disorders, leprosy,	[29,30]
<i>C. adenogonium</i> Stend. Ex A. Rich.	Leaves	wounds, female sterility, etc. Liver and gallbladder diseases, colds, bronchitis, malaria, anemia, migraine, wounds, amoebic dysentery, diarrhea, urinary disorders, bilious and hematuric fevers, edema, albuminuria, anorexia, cough, severe jaundice, Hypertension, hepatitis, epilepsy, snakebites, asthenia.	[29]
	Bark	Epigastralgia, intestinal parasites, sexual weakness, vomiting, analgesic, aphrodisiac, anthelmintic.	
	Roots	Diseases of the stomach, cough, intestinal parasites, syncope, snake bite, pregnancy bleeding.	
C. apiculatum Sond.	Bark	Aphrodisiac	[31]
C. camporum Engl.	Bark, roots	Cancer	[14]
<i>C. coccineum</i> (Sonn.) Lam.	Leaves	Herbal tea cholagogue	[32]
	Roots and fruits	Anthelmintic	
C. collinum Fresen	Roots	Painful legs, cramps, and joint pains.	[33]
C. crotonoides Hutch et Dalz	Leafy stems	Pneumonia, bronchitis, liver disorders, cholagogue, general fatigue, rheumatism	[29]
<i>C. erythrophyllum</i> (Burch.) Sond.	Leaves, roots, bark	Abdominal pain, sexually transmitted diseases, diarrhea, dysentery, coughs, colds, infertility, sores, and wounds.	[34,35]
<i>C. fragrans</i> F. Hoffm.	Whole plant	Leprosy, coughs, diarrhea, pain and inflammation, jaundice, ulcers, wounds, and cancers.	[12,15,32,36]
	Leaves	Dysentery, burns, and wounds	
	Leafy twigs	Diarrhea,	
	Fruits	Wounds	
	Roots	Diarrhea, pain	10.01
<i>C. glutinosum</i> Perr. ex DC	Leaves	Cholagogues, depurative, diuretic, pectoral, malaria.	[32]
	Leafy twigs	Cholagogues, anemia, childhood gastritis, jaundice, edema, malaria, and various eye conditions.	
	Powdered fruit	Wounds, syphilis.	
C. hartmannianum (Schweinf)	Wood, bark	Febrile, jaundice, bacterial infections, cough, tuberculosis, neoplasia	[37-39]
C. hereroense Schinz	Leaves	Abdominal pain, sexually transmitted diseases	[40]
	Pooto stom	Menstruation, stomachache	[31,33]
<i>C. imberbe</i> Engl. & Diels	Roots, stem, leaves		[01,00]

Table 1. Traditional uses of some *Combretum* species reported in the literature

Species	Used parts	Traditional uses	References
C. leprosum Mart.	Leaves and flowers	Inflammation, pain, treatment of wounds, sedative, diarrhea,	[41,42]
		expectorant, and antitussive.	
<i>C. micranthum</i> G. Don	Leaves	Diarrhea, dysentery, colic, stomach disorder, digestion disorder, malaria,	[14,29,32,43]
		beriberi, intestinal parasites, asthenia, cough, bronchitis, fevers, lumbago,	
		gallbladder disease, anemia, gonorrhea,	
		hypertension, mastitis, bilious and	
		hematuric fevers, cancer.	
	Roots	Female sterility, intestinal parasites, syphilis, enuresis, stimulant,	
		constipation, fever gastritis, jaundice,	
		trichocephalosis, cancer.	
	Bark	Contusions, sprains, massage before	
		and after muscular effort, lumbago,	
	Fruits	cancer. Hypertension, stomatitis, hypotension	
	Seeds	Candidiasis, thrush, boils abscess, leucorrhea.	
C. molle G. Don	Leaves, roots,	Gonorrhoea, syphilis, influenza, edema,	[14,35,40]
	bark	skin diseases and wounds, diarrhea,	• • • •
		stomachache, measles, cough, eye	
		pain, asthma, and cancer.	
<i>C. mucronatum</i> Schumach. & Thonn.	Stem bark	Anthelmintic	[32]
	Leaves	Wounds and injuries.	
<i>C. nigricans</i> Lepr. ex Guill. et Perr.	Leafy stems	Hematuria, gastric diseases, malaria, fever, strengthening babies, cancer	[22,28]
	Bark	Headache, intestinal disorders, rheumatism	[30]
	Roots	Mental illness	
C. nioroense Aubrev.	Leafy stems	Malaria, dysentery, diarrhea,	[44]
ex keay	,	strengthening babies	
	Leaves	Metastasis, neck, head, face and breast cancers	[43]
C. paniculatum Vent.	Gall	Vomit	[28]
·	Leaves	Gastric colic, hernial pain,	
		strengthening, rickets, cough, injuries,	
		hemorrhoids, wounds	
	Roots	Diarrhea, malaria with inflammation of the spleen, wounds	
C. psidioides Welw.	Roots, leaves	Diarrhea, muscle pain, edema.	[12]
C. racemosum P.	Whole plant	Anthelmintic, gastrointestinal and	[32,45,46]
Beauv.		genito-urinary infections, hemorrhoid,	
		convulsive coughing, tuberculosis,	
	Leaves,	toothache, and male sterility Hypertension	
	rootbark	пурецензіон	
C. sericeum G. Don	Leafy stems	Malaria, weight delay, open fractures,	[28,47]
	-	stomach ache, conjunctivitis, fevers.	
	Roots	Diarrhea, pneumonia, wounds.	
<i>C. tomentosum</i> G. Don	Bark, leaves	Breast, neck, throat and head Cancers	[43]
C. zeyheri Sond.	Roots, leaves	Diarrhea, cancer	[12]

The cytotoxic effect of methanol (80%) extract from the root of Combretum adenogonium has been investigated, and authors found that the extract inhibits human prostate cancer cell lines (PC-3) with an IC₅₀ of 24 μ g/mL [48]. The methanol extract from leaves of Combretum fragrans inhibited several human cancer cell lines such as T24 (Bladder cancer), Hela (Cervical cancer), and MCF-7 (Breast cancer) with IC₅₀ around 4.3 µg/mL for both T24 and Hela cells and 14.6 µg/mL for MCF-7 cells [13]. A study has highlighted the cytotoxicity of dichloromethane extract from leaves of Combretum fruticosum on human cancer cells such as adenocarcinoma human alveolar basal epithelial cells (A549), glioblastoma cells (U373; Hs683), human prostate cancer cells (PC3) and Brain primary metastases cells (Kaka) with IC₅₀ of 9 µg/mL, 7 µg/mL, 8.5 µg/mL, 10 µg/mL and 8.3 µg/mL respectively [49]. The trachelogenin compound isolated from the talks of this plant has demonstrated cytotoxic properties on human brain cancer cells (SF-295) and human leukemia cells (HL-60). This compound also presented antiproliferative activity on human colon cancer cells (HCT-116) with an IC₅₀ value of 4.8 μ M [50]. has showed been that Combretum It microphyllum has potential antimutagenic activity and protective effects against cancer. Indeed, the crude extract of this plant inhibits the genotoxic end-points induced by 4-nitroguinoline 1-oxide (4NQO). mitomycin-C (MMC), and ethvl methanesulfonate (EMS) in vitro. Isolated compounds n-tetracosanol, eicosanoic acid, and arjunolic acid, presented an antimutagenicity of 42 ± 9.6%, 36 ± 1.5%, and 44 ± 0.18% in S. typhimurium TA98 [51]. Ethanol (80%) extract of Combretum paniculatum has presented a potent cytotoxic effect on Hela cancer cells at 500 µg/mL. The extract inhibited almost 70% of the growth of this cancer cell line [52]. Jurkat cells exposed to Combretum platypetalum extract at 31,25 µg/ml during 72 hours showed a significant reduction of cell viability of this cell line [53]. Ethanol extract from leaves of Combretum quadrangulare presented a cytotoxic effect on A549 cancer cells after 48 hours of treatment with an IC₅₀ of 136.1 μ g/mL [54]. The ethyl acetate extract from leaves of Combretum rupicola showed significant anticancer activities against four cell lines and the most significant activity was observed against MCF-7 cells with an IC_{50} of 65.9 $\mu g/mL$ [55]. All these results contribute to highlighting the anticancer potential of Combretum species and in the same way justify their uses in traditional medicine for cancer treatment.

3.3 Potential Anticancer Compounds Isolated from *Combretum* Species

Many potential anticancer compounds have been isolated from combretum species. The cytotoxicity important of some isolated compounds is presented in Table 2 and their structures are shown in Fig. 1. Few of these compounds have been investigated in deep for preclinical studies. The myricitrin isolated from C. lanceolatum proved to be most effective in inhibiting the growth of leukemic HL-60 cells. The mechanism of this inhibition is through the inhibition of the topoisomerase IIa [56]. The trachelogenin compound isolated from C. fruticosum induced autophagy cell death in human colon cancer cells (HCT-116) with LC3 activation and altered the expression levels of Beclin-1 [50]. The triterpenoid (1-O-[α-L-(rhamnopyranosyl]-23-acetoxy-3_β-acetylimberbic acid) isolated from C. Sundaicum inhibited KB, MCF7, and HCT-116 cells growth by binding to Bcl-xL protein [57]. The pentacyclic triterpene (3β, 6β, 16β-trihydroxylup-20(29)-ene) isolated from C. leprosum induced apoptosis in MCF7 cells by increasing levels of both cleaved caspase-9 and intracellular ROS [58].

Among isolated compounds from Combretum species, Combretastatins were the most subject to research. They are first isolated from the South African Combretum caffrum specie [59]. Other Combretum species also contain these molecules [60,61]. Combretastatins are simple compounds formed of two phenyl rings linked by a chain of two carbon atoms. They consist of several series belonging to the stilbenoid class of compounds [62]. Several studies have been undertaken to highlight the anticancer properties of these compounds [63-65]. Combretastatins belonging to the A and B series present potent anticancer activity with the combretastatin A4 molecule being the most potent due to its broadspectrum cytotoxicity against a variety of tumor cells [66]. It has been demonstrated that Combretastatin A4 can induce cell cycle arrest at M-phage and apoptosis in various cancer cells through a mechanism of action involving inhibition of tubulin polymerization [64,65]. It has also been revealed that combretastatin A4 can reverse daunorubicin resistance acquisition in the P-388 cell line [67]. This proves that the molecule could play an important role in the treatment of certain resistant forms of tumors. The cis isomer of this molecule is the active form while the trans isomer is inactive [63]. The cis

isomer can spontaneously isomerize into the trans isomer. This problem of isomerism added to its low solubility and its poor bioavailability has led to the development of analogs such as combretastatin A4 phosphate which is water-

soluble, bioavailable, and biologically active [68]. Combretastatin A4 phosphate undergoes clinical trials as an anticancer agent for the treatment of solid cancers [69,70].

Table 2. Cytotoxicity (IC ₅₀) of isolated compounds from <i>Combretum</i> species on some cancer
cell models

Species	Plant part	Compounds	Cell lines	Histotype	IC ₅₀	References
<i>Combretum caffrum</i> (Eckl. and Zeyh.) Kuntze	Twigs and leaves	(1) Combretastatin A4	P-388	murine lymphocytic leukemia	0.011 µg/mL	[59]
<i>Combretum</i> Leaves fragrans F. Hoffm.	Leaves	(2) Combretin A	MCF-7	Breast adenocarcinoma	16.1 μg/mL	[15,36]
			U87	Glioblastoma	29.3 µM	
			PC-3	Prostate adenocarcinoma	22.57 µM	
		(3) Combretin B	MCF-7	Breast adenocarcinoma	< 1 µg/mL	[36]
		(4) Cirsilineol	U87	Glioblastoma	65.46 µM	[15]
			PC-3	Prostate adenocarcinoma	30.46 µM	
Combretum racemosum P. Beauv.	Roots	(5) 3-O-β-acetyl- ursolic acid	HL-60	Leukemia	14.5 µM	[71]
			K562	Leukemia	50.0 µM	
		(6) Betulinic acid	HL-60	Leukemia	35 µM	[71]
			K562	Leukemia	> 50 µM	
		(7) Quadranoside II	HL-60	Leukemia	13 µM	[71]
			K562	Leukemia	44.0 µM	
<i>Combretum</i> stems <i>griffithii</i> Heurck & Müll.	stems	(8) 1-(5-hydroxy-2,4- dimethoxyphenyl)-3- (4-hydroxy-3- methoxyphenyl)prop ane	KB	Cervical adenocarcinoma	24.63 μΜ	[72]
			MCF-7	Breast adenocarcinoma	23.12 μΜ	
			NCI-H18	Small cell lung cancer	34.65 μΜ	
		(9) (2S)-3′,4′- dihydroxy-5,7- dimethoxyflavan	KB (Hela derivativ e)	Cervical cancer	31.82 μΜ	[72]
			MCF-7	Breast adenocarcinoma	49.15 μΜ	
			NCI-H18	Small cell lung cancer	10.41 μΜ	
Combretum Ieprosum Mart.	Flowers	(10) 3β,6β,16β- Trihydroxylup- 20(29)-ene	MCF-7	Breast adenocarcinoma	1.36 µg/mL	[58]
** *			HepG2	Hepatoma	6.50 µg/mL	

Species	Plant part	Compounds	Cell lines	Histotype	IC ₅₀	References
			T24	Bladder carcinoma	5.55 μg/mL	
			HCT-116	Colorectal carcinoma	7.05 μg/mL	
			HT-29	Colorectal adenocarcinoma	8.0 μg/mL	
			CACO-2	Colorectal adenocarcinoma	8.55 μg/mL	
Combretum quadrangula re Kurz	Leaves	(11) Methyl quadrangularate D	Colon 26-L5	Murine colon cancer	5.42 µM	[73]
			HT-1080	Fibrosarcoma	40.0 µM	
			HeLa	Cervical carcinoma	50.9 µM	
			A-549	Lung adenocarcinoma	63.5 µM	
		(12) Methyl quadrangularate B	Colon 26-L5	Murine colon cancer	9.54 µM	[73]
			HT-1080	Fibrosarcoma	53.3 µM	
			HeLa	Cervical carcinoma	56.0 µM	
			A-549	Lung adenocarcinoma	>100.0 µM	
		(13) Combretic acid C	K562	Leukemia	9.7 µM	[74]
		(14) Kamatakenin	Colon 26-L5	Murine colon cancer	3.0 µM	[75]
			HT-1080	Fibrosarcoma	3.2 µM	
			HeLa	Cervical carcinoma	10.6 µM	
			A-549	Lung adenocarcinoma	40.1 µM	
		(15) Isokaempferide	Colon 26-L5	Murine colon cancer	4.5 µM	[75]
			HT-1080	Fibrosarcoma	0.8 µM	
			HeLa	Cervical carcinoma	8.2 µM	
			A-549	Lung adenocarcinoma	39.2 µM	
		(16) 5,7,4'- trihydroxy-3,3'- dimethoxyflavone	Colon 26-L5	Murine colon cancer	1.8 µM	[75]
			HT-1080	Fibrosarcoma	1.5 µM	
			HeLa	Cervical cancer	6.8 µM	
			A-549	Lung adenocarcinoma	95.3 µM	
		(17) 5,4'-dihydroxy- 3,7,3'- trimethoxyflavone	Colon 26-L5	Murine colon cancer	5.2 µM	[75]
			HT-1080	Fibrosarcoma	3.5 µM	
			HeLa	Cervical carcinoma	38.4 µM	

Species	Plant part	Compounds	Cell lines	Histotype	IC ₅₀	References
			A-549	Lung adenocarcinoma	41.9 µM	
Combretum Le nigricans Lepr. ex Guill. et Perr.	Leaves	(18) 20,24-epoxy- 11α,25-dihydroxy- dammar-3-one	U-373	Glioblastoma	31.6 µg/mL	[76]
			HCT-15	Colon adenocarcinoma	30.4 µg/mL	
			A-549	Lung adenocarcinoma	29.7 µg/mL	
			J82	Bladder carcinoma	29.6 µg/mL	
		(19) 11α-acetoxy- 20,24-epoxy-25- hydroxy-dammar-3- one	U-373	Glioblastoma	51.7 μg/mL	[76]
			HCT-15	Colon adenocarcinoma	88.4 µg/mL	
			A549	Lung adenocarcinoma	>100 µg/mL	
			J82	Bladder carcinoma	80.6 µg/mL	
Combretum Le oliviforme Chao	Leaves	(20) 23-acetoxy- 3β-acetylimberbic acid-29-methyl ester	SPC-A1	Lung cancer	1.09 µM	[77]
			SGC- 7901	Gastric cancer	0.69 µM	
			K562	Leukemia	2.10 µM	
		(21) 23-O-[α-L- rhamnopyranosyl]- 1,3β- diacetylimberbic acid	SPC-A1	Lung cancer	25.20 μΜ	[77]
			SGC- 7901	Gastric carcinoma	5.44 µM	
			K562	Leukemia	2.38 µM	
Combretum Sta fruticosum (Loefl.) Stuntz	Stalks	(22) (-)- Trachelogenin	HL-60	Leukemia	32.4 µM	[50]
			OVCAR- 8	Ovarian carcinoma	3.5 µM	
			HCT-116	Colorectal carcinoma	1.9 µM	
			HCT-8	lleocecal adenocarcinoma	5.2 µM	
			PC-3	Prostate cancer	15 µM	
			SF295	Glioblastoma	0.8 µM	
Combretum sundaicum Miq.	Leaves and flowers	(23) 1-o-[α-L- rhamnopyranosyl]- 23-acetoxy-3β- acetylimberbic acid- 29-methyl ester	КВ	Cervical adenocarcinoma	0.9 µM	[57]

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Species	Plant part	Compounds	Cell lines	Histotype	IC ₅₀	References
			HCT-116	Colorectal carcinoma	0.3 µM	
			MCF-7	Breast adenocarcinoma	0.6 µM	
Combretum L platypetalum Welw. ex M.A. Lawson	Leaves	(24) 3-o-(β-D- glucopyranosyl)- 5,7,3',4',5'- pentahydroxyflavone	Jurkat T	Leukemia	3.98 µg/mL	[78]
			HL-60	Leukemia	14.18 µg/mL	
		(25) 3-o-(α-L- rhamnopyranosyl)- 5,7,3',4',5'- pentahydroxyflavone	Jurkat T	Leukemia	19.33 µg/mL	[78]
			HL-60	Leukemia	28.7 µg/mL	
Combretum Lanceolatum Pohl.	Leaves and branches	(26) myricetin	MCF7	Breast adenocarcinoma	158.4 μΜ	[56]
			PC-3	Prostate cancer	182.8 μΜ	
			HT-29	Colon adenocarcinoma	95 µM	
			786-0	Kidney carcinoma	124.6 μΜ	
			HL-60	Leukemia	53.4 µM	
Combretum Roo laxum Jacq.	Roots	(27) 4'-hydroxy- 3,3',4-trimethoxy-5- (3,4,5- trimethoxyphenoxy)- bibenzyl	MCF-7	Breast adenocarcinoma	72.69 μΜ	[79]
			NCI/ADR -RES	Ovary carcinoma, multidrug-resistant	32.09 µM	
		(28) 2,7-dihydroxy- 4,6- dimethoxyphenanthr ene	786-0	Kidney carcinoma	73.26 μΜ	[79]
			NCI/ADR -RES	Ovary carcinoma, multidrug-resistant	83.99 μΜ	
		(29) 2,6-dihydroxy- 3,4,7- trimethoxyphenanthr ene	786-0	Kidney carcinoma	64.27 μΜ	[79]
		(30) 6- methoxycoelonin	MCF-7	Breast adenocarcinoma	46.99 μΜ	[79]
			UACC- 62	Human melanoma	2.59 µM	
			NCI/ADR -RES	Ovary carcinoma, multidrug-resistant	58.83 μΜ	
			786-0	Kidney carcinoma	56.98 μΜ	
		(31) 2,6-dihydroxy- 3,4,7-trimethoxy-	MCF-7	Breast adenocarcinoma	42.01 μΜ	[79]

Plant part	Compounds	Cell lines	Histotype	IC ₅₀	References
	9,10- dihvdrophenanthrene				
Leaves	(32) Combretaside G	NCI- H460	Lung carcinoma	3.9 µM	[80]
Roots and Leaves	(33) Tetrahydrofuran lignan	HT-29	Colon adenocarcinoma	3.9 µM	[81]
	(34) 2,3-seco-lup- 20(29)-en-2,3-dioic acid	786-0	Kidney carcinoma	0.5 µM	[81]
		HT-29	Colon adenocarcinoma	2.9 µM	
	CH ₃ 2	R = OH	$H_{3}CO$ $H_{3}CO$ $H_{3}CO$ OH yranoside		CH₃) OH
	ро н		соон но _{м,} но нон ₂ с ^{чч}		
ОН ОСН ₃ 8	OH H ₃ CO OCH ₃	осн ₃ 9	ОН	ОН 10	Сон
	Но но	OHC ₄	Коон		Коон
	Leaves Roots and Leaves f(x) = 1 f(x) = 1 f	$\begin{array}{c} 9,10-\\ dihydrophenanthrene\\ Leaves (32) Combretaside G\\ \hline Roots and Leaves (33) Tetrahydrofuran lignan \\ (34) 2,3-seco-lup-20(29)-en-2,3-dioic acid \\ \hline (34) 2,3-seco-lup-20(29)-en-2,3-dioic acid \\ \hline (34) 2,3-seco-lup-20(29)-en-2,3-dioic acid \\ \hline (54) (54) (54) (54) (54) (54) (54) (54)$	$\begin{array}{c} \begin{array}{c} \begin{array}{c} \begin{array}{c} 1 \\ \begin{array}{c} 0 \\ \end{array} \\ \end{array} \\ \begin{array}{c} 0 \\ \end{array} \\ \end{array} \\ \begin{array}{c} 0 \\ \end{array} \\ \end{array} \\ \begin{array}{c} 0 \\ \end{array} \\ \begin{array}{c} 0 \\ \end{array} \\ \end{array} \\ \begin{array}{c} 0 \\ \end{array} \\ \begin{array}{c} 0 \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} 0 \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} 0 \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} 0 \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} 0 \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} 0 \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} 0 \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} 0 \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} 0 \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} 0 \\ \end{array} \\ \begin{array}{c} 0 \\ \end{array} \\ \begin{array}{c} 0 \\ \end{array} \\$	$\begin{array}{c c} & & & & & & & & & & & & & & & & & & &$	$\begin{array}{c c c c c } \hline lines \\ \hline lines \\ \hline 0,10^{-} \\ \hline 1 \\ \hline 0,10^{-} \\ \hline 0,10^{-} \\ \hline 1 \\ 1 \\$

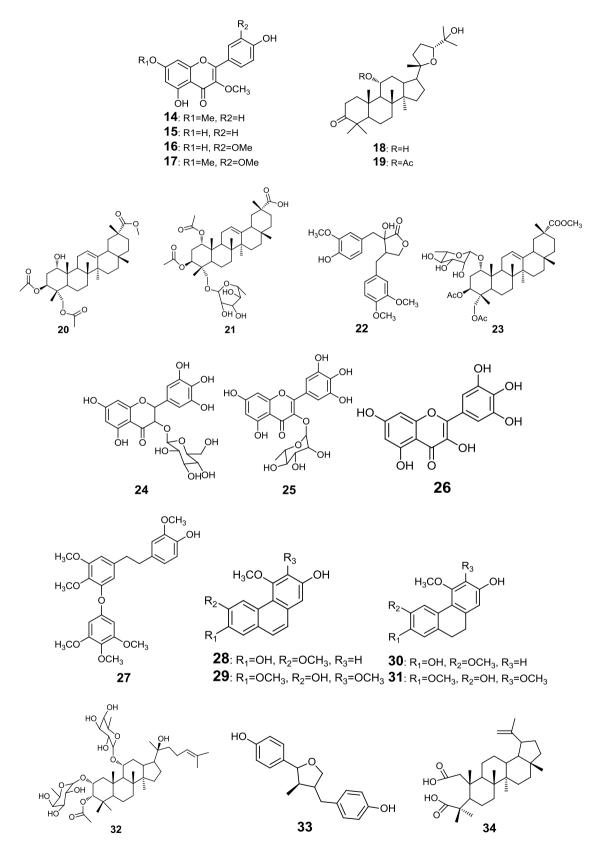


Fig. 1. Chemical structures of some important cytotoxic isolated compounds from *Combretum* species

The different numbers (1-34) correspond to the compound names in Table 2

4. CONCLUSION AND FUTURE PERSPECTIVES

In this review, we discussed the traditional uses of Combretum species reported in some ethnobotanical studies over the world with special emphasis on their anticancer uses. We also highlighted several pharmacological studies realized on Combretum species which revealed their cytotoxicity activities on various cancer cell models pointing out the anticancer potential of these plant species. Finally, some potential anticancer compounds isolated from Combretum species were reported as well. This proves that Combretum species could be an important resource for researchers to look for new anticancer molecules that may contribute to the fiaht against cancers. In perspectives. ethnobotanical studies must be undertaken on Combretum species especially the 12 Combretum species present in Burkina Faso to highlight their anticancer uses and correlate this finding to clinical efficacy which will be evaluated by controlled clinical studies.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

COMPETING INTERESTS

Author has declared that no competing interests exist.

REFERENCES

- Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. CA Cancer J Clin. 2011; 61(2):69-90. DOI: 10.3322/caac.20107
- Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. CA Cancer J Clin. 2015; 65(2):87-108.

DOI: 10.3322/caac.21262.

 Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2018;68(6):394-424. DOI: 10.3322/caac.21492

 Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global Cancer Statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2021;71(3): 209-249.

DOI: 10.3322/caac.21660

 GLOBOCAN. Burkina Faso; 2020. Available:https://gco.iarc.fr/today/data/facts heets/populations/854-burkina-faso-factsheets.pdf Access on 15 Dec. 2022)

 Beiersmann C, Sanou A, Wladarsch E, De Allegri M, Kouyaté B, Müller O. Malaria in rural Burkina Faso: Local illness concepts, patterns of traditional treatment and influence on health-seeking behaviour. Malar J. 2007;6:106.

DOI: 10.1186/1475-2875-6-106.
7. Nadembega P, Boussim JI, Nikiema JB, Poli F, Antognoni F. Medicinal plants in Baskoure, Kourittenga Province, Burkina Faso: An ethnobotanical study. J Ethnopharmacol. 2011;133(2):378-395. DOI: 10.1016/j.jep.2010.10.010.

 Jansen O, Angenot L, Tits M, Nicolas JP, De Mol P, Nikiéma J-B, Frédérich M. Evaluation of 13 selected medicinal plants from Burkina Faso for their antiplasmodial properties. J Ethnopharmacol. 2010;130: 143-150.

DOI: 10.1016/j.jep.2010.04.032

9. Sofowora A, Ogunbodede E, Onayade A. The role and place of medicinal plants in the strategies for disease prevention. Afr J Tradit Complement Altern Med. 2013;10 (5):210-229.

DOI: 10.4314/ajtcam.v10i5.2.

 Dehelean CA, Marcovici I, Soica C, Mioc M, Coricovac D, Iurciuc S, et al. Plantderived anticancer compounds as new perspectives in drug discovery and alternative therapy. Molecules. 2021;26(4): 1109.

Available:https://doi.org/10.3390/molecules 26041109

- Lichota A, Gwozdzinski K. Anticancer Activity of natural compounds from plant and marine environment. Int J Mol Sci. 2018;19(11):3533. DOI: 10.3390/ijms19113533.
- 12. Fyhrquist P, Mwasumbi L, Haeggström CA, Vuorela H, Hiltunen R, Vuorela P. Ethnobotanical and antimicrobial investigation on some species of

Terminalia and *Combretum* (Combretaceae) growing in Tanzania. J Ethnopharmacol. 2002;79(2):169-177. DOI: 10.1016/s0378-8741(01)00375-0

 Fyhrquist P, Mwasumbi L, Vuorela P, Vuorela H, Hiltunen R, Murphy C et al. Preliminary antiproliferative effects of some species of Terminalia, Combretum and Pteleopsis collected in Tanzania on some human cancer cell lines. Fitoterapia. 2006; 77(5):358-366.

DOI: 10.1016/j.fitote.2006.05.017

 Malami I, Jagaba NM, Abubakar IB, Muhammad A, Alhassan AM, Waziri PM et al. Integration of medicinal plants into the traditional system of medicine for the treatment of cancer in Sokoto State, Nigeria. Heliyon. 2020;6(9):e04830.

DOI: 10.1016/j.heliyon.2020.e04830.

15. Gade IS, Chadeneau C, Simo RT, Talla E, Atchade ADT, Seité P et al. A new phenyl alkyl ester and a new combretin triterpene derivative from *Combretum fragrans* F. Hoffm (Combretaceae) and antiproliferative activity. Open Chemistry. 2020;18(1):1523-1531.

DOI:10.1515/chem-2020-0167

- Coulidiati TH, Millogo-Koné H, Lamien-Meda A, Yougbaré-Ziébrou M, Millogo-Rasolodimby J, Nacoulma OG. Antioxidant and antibacterial activities of two *Combretum* species from Burkina Faso. Res J Med Plants. 2011;5:42-53.
- Saotoing 17. Dawe Α, Ρ, Tsala DE, Phytochemical Habtemariam S. Combretum constituents of loefl. (Combretaceae). Pharmaceutical Crops. 2013:4:38-59.
- Bantho S, Naidoo Y, Dewir YH, Bantho A, Murthy HN. Chemical composition of *Combretum erythrophyllum* leaf and stem bark extracts. Horticulturae. 2022;8:755. Available:https://doi.org/10.3390/ horticulturae8080755
- Moyo M, Aremu AO, Van Staden J. Medicinal plants: An invaluable, dwindling resource in Sub-Saharan Africa. J Ethnopharmacol. 2015;174:595-606.
 DOI: 10.1016/j.jep.2015.04.034.
- Abdullahi AA. Trends and challenges of traditional medicine in Africa. Afr J Tradit Complement Altern Med. 2011;8:115-123. DOI: 10.4314/ajtcam.v8i5S.5

- Benoît É. Les changements climatiques : vulnérabilité, impacts et adaptation dans le monde de la médecine traditionnelle au Burkina Faso. Vertigo 2008;8(1).
 DOI : 10.4000/vertigo.1467.
- 22. Thiombiano A. Les Combretaceae du Burkina Faso: Taxonomie, écologie, dynamique et régénération des espèces. Thèse de doctorat d'État, Université de Ouagadougou, Burkina Faso. 2005:290.
- Thiombiano A, Schmidt M, Kreft H, Guinko S. Influence du gradient climatique sur la distribution des espèces de Combretaceae au Burkina Faso (Afrique de l'Ouest). Candollea. 2006;61:189-213.
- Shalukoma C, Bogaert J, Duez P, Stévigny 24. C, Pongombo C, Visser M. Les plantes medicinales de la region montagneuse de Kahuzi-Biega en République Democratique du Congo: Utilisation. accessibilité et consensus des tradipraticiens. Bois et Forêts des Tropiques. 2015;326:43-55.
- Schultz F, Anywar G, Wack B, Quave CL, Garbe LA. Ethnobotanical study of selected medicinal plants traditionally used in the rural Greater Mpigi region of Uganda. J Ethnopharmacol. 2020;256:112742.

DOI: 10.1016/j.jep.2020.112742

 Lingani A, Lompo LF, Guissou IP, Nikiema JB. Médecine traditionnelle et maladies des reins au Burkina Faso [Traditional medicine in kidney diseases in Burkina Faso]. Nephrol Ther. 2010;6(1): 35-39.

DOI: 10.1016/j.nephro.2009.07.011.

- Ouôba P, Lykke AM, Boussim J, Guinko S. La flore médicinale de la forêt classée de Niangoloko (Burkina Faso). Etudes flor vég Burkina Faso. 2006;10:5-16.
- Zerbo P, Millogo-Rasolodimby J, Nacoulma OG, Van Damme P. Plantes médicinales et pratiques médicales au Burkina Faso: Cas des Sanan. Bois et Forêts des Tropiques 2011;307(1):41-53.
- 29. Nacoulma OG. Plantes médicinales et pratiques médicales traditionnelles au Burkina Faso: cas du plateau central. Thèse de doctorat d'État, Faculté des sciences et techniques, Université de Ouagadougou, Burkina Faso, tome 1, 320 p., tome 2, 285 p; 1996.

- Arbonnier M. Arbres, arbustes et lianes des Zones sèches d'Afrique de l'Ouest. Ed. ISBN CIRAD, Pont-sur-Yonne, 2002;392:574.
- Bruschi P, Morganti M, Mancini M, Signorini MA. Traditional healers and laypeople: a qualitative and quantitative approach to local knowledge on medicinal plants in muda (Mozambique). J Ethnopharmacol. 2011;138:543-563.
- 32. Boulard B. Plantes médicinales du monde: Croyances et réalités. Ed. ESTEM, Paris. 2001:660.

ISBN 2843711177

- Tshikalange TE, Mophuting BC, Mahore J, Winterboer S, Lall N. An Ethnobotanical study of medicinal plants used in villages under Jongilanga tribal council, Mpumalanga, South Africa. Afr J Tradit Complement Altern Med. 2016:13(6):83-89.
- 34. Martini ND, Katerere DRP, Eloff JN. Biological activity of five antibacterial flavonoids from *Combretum erythrophyllum* (Combretaceae). J Ethnopharmacol. 2004; 93:207-212.
- 35. Cock IE, Van Vuuren SF. A comparison of the antimicrobial and toxicity of six *Combretum* and two *Terminalia* species from southern Africa. Pharmacognosy Magazine. 2015:11(41): 208-218.
- Mbiantcha M, Almas J, Dawe A, Faheem A, Sidra Z. Analgesic, anti-inflammatory and anticancer activities of Combretin A and Combretin B isolated from Combretum fragrans F. HOFFM (Combretaceae) leaves. Inflammopharmacology. 2018;26 (6):1429-1440.

DOI: 10.1007/s10787-017-0421-5.

 Mohieldin EAM, Muddathir AM, Mitsunaga T. Inhibitiry activities of Sudanese medicinal plants on Porphyromonas gingivalis and matrix metalloproteinase-9 and isolation of bioactive compounds from *Combretum hartmannianum* (Schweinf) bark. BMC Complement Altern Med. 2017; 17(1):224.

DOI: 10.1186/s12906-017-1735-y.

 Hassan LE, Ahamed MB, Majid AS, Baharetha HM, Muslim NS, Nassar ZD, et al. Correlation of antiangiogenic, antioxidant and cytotoxic activities of some Sudanese medicinal plants with phenolic and flavonoid contents. BMC Complement Altern Med. 2014;14:406.

DOI: 10.1186/1472-6882-14-406.

39. Salih EYA, Julkunen-Tiitto R, Luukkanen O, Fahmi MKM, Fyrquist P. Hydrolyzable (ellagitannins), flavonoids, tannins triterpenes pentacyclic and their glycosides in antimycobacterial extracts of ethnopharmacologically the selected Sudanese medicinal plant Combretum hartmannianum Schweinf. Biomed Pharmacother, 2021:144:112264.

DOI: 10.1016/j.biopha.2021.112264.

- 40. Martini ND, Eloff JN. The preliminary isolation of several antibacterial compounds from *Combretum erythrophyllum* (Combretaceae). J Ethnopharmacol. 1998;62:255-263.
- Horinouchi CD, Mendes DA, Soley Bda S, 41. Pietrovski EF, Facundo VA, Santos AR et Combretum leprosum Mart. al. (Combretaceae): Potential as an anti-inflammatory antiproliferative and agent. .1 Ethnopharmacol. 2013;145(1):311-319.

DOI: 10.1016/j.jep.2012.10.064.

- Lacouth-Silva F, Xavier CV, Sutúbal SDS, Pontes AS, Nery NM, de Castro OB et al. The effect of 3β, 6β, 16β-trihydroxylup-20(29)-ene lupine compound isolated from *Combretum leprosum* Mart. On peripheral blood mononuclear cells. BMC Complement Altern Med. 2015;15(1):420. DOI: 10.1186/s12906-015-0948-1.
- 43. Abubakar IB, Ukwuani-Kwaja AN, Garba AD, Singh D, Malami I, Salihu TS et al. Ethnobotanical study of medicinal plants used for cancer treatment in Kebbi state, North-west Nigeria. Acta Ecologica Sinica. 2020;40(4):306-314.
- Coulidiati TH, Millogo-Koné H, Lamien-44. Meda A, Lamien CE, Lompo Μ, Kiendrébéogo M et al. Antioxidant and antibacterial activities of Combretum Aubrév. nioroense Ex Keav (Combretaceae). Pak J Biol Sci. 2009;12(3):264-269.

DOI: 10.3923/pjbs.2009.264.269.

 Okwuosa C, Unekwe P, Nwobobo E, Chilaka K. The anti-ulcer activities of leaf extracts of *Combretum racemosum* (family: combretaceae). Journal of Biomedical Investigation. 2006;4(1):9-14. DOI: 10.4314/jbi.v4i1.30408

- Manga FN, Khattabi CE, Fontaine J, Berkenboom G, Duez P, Nzunzu JL et al. Vascular effects and antioxidant activity of two *Combretum* species from Democratic Republic of Congo. J Ethnopharmacol. 2012;142(1):194-200.
- 47. Sini JM, Umar IA, Anigo KM, Stancheva I, Bage EN, Mohammed R. Antidiarrhoeal activity of aqueous extract of *Combretum sericeum* roots in rats. Afr J Biotech. 2008;7(17):3134-3137.
- 48. Elnour MA, Penech F, Ibrahim S, Khalid A. *Combretum adenogonium* induce anticancer and antioxidant effects in prostate cancer cell line. Significances Bioengineering & Biosciences 2018;2(2): 138-140.

DOI: 10.31031/SBB.2018.02.000534

- 49. Balde ES, Camara AK, Traoré MS, Baldé NM, Megalizzi V, Pieters L et al. The hypoglycemic and cytotoxic activity of the leave extract of *Combretum glutinosum* Perr ex DC. J Pharmacogn Phytochem. 2019;8(4):2230-2237.
- 50. Moura AF, Lima KSB, Sousa TS, Marinho-Filho JDB, Pessoa C, Silveira ER et al. *In vitro* antitumor effect of a lignan isolated from *Combretum fruticosum*, trachelogenin, in HCT-116 human colon cancer cells. Toxicol *In Vitro*. 2018;47:129-136.

DOI: 10.1016/j.tiv.2017.11.014.

51. Makhafola TJ, Elgorashi EE, McGaw LJ, Awouafack MD, Verschaeve L, Eloff JN. Isolation and characterization of the compounds responsible for the antimutagenic activity of Combretum microphyllum (Combretaceae) leaf extracts. BMC Complement Altern Med. 2017:17(1):446.

DOI: 10.1186/s12906-017-1935-5.

 Sowemimo A, van de Venter M, Baatjies L, Koekemoer T. Cytotoxic activity of selected Nigerian plants. Afr J Tradit Complement Altern Med. 2009;6(4):526-528.
 DOI: 10.4314/aitcam.v6i4.57186

DOI: 10.4314/ajtcam.v6i4.57186

53. Chiramba O, Mukanganyama S. Cytotoxic effects of *Combretum platypetalum* Welw. ex M.A. Lawson subsp. oatesii (Rolfe) Exell (Combretaceae) leaf extracts on Jurkat T-cells and reversal of effects by reduced glutathione. Journal of Biologically Active Products from Nature. 2016;6(3): 250-265.

DOI: 10.1080/22311866.2016.1232626.

- 54. Chittasupho C. Athikomkulchai S. Nanoparticles of Combretum quadrangulare leaf extract induce cytotoxicity, apoptosis, cell cycle arrest and anti-migration in lung cancer cells. Journal of Drug Delivery Science and Technology 2018:45:378-387.
- Santos SN, Kavamura V, Castanha RF, 55. Andreote FD. Carvalho JE. Queiroz SCN Antitumoral. antioxidant et al. and antimicrobial molecules from Combretum rupicola. Int J Pharma Bio Sci. 2013;4(1):422-428.
- 56. Perdomo RT, Defende CP, da Silva Mirowski P, Freire TV, Weber SS, Garcez WS et al. Myricitrin from *Combretum lanceolatum* exhibits inhibitory effect on DNA-topoisomerase type IIα and protective effect against in vivo doxorubicin-induced mutagenicity. J Med Food. 2021;24(3):273-281.

DOI: 10.1089/jmf.2020.0033

57. Litaudon M, Jolly C, Le Callonec C, Cuong DD, Retailleau P, Nosjean O et al. Cytotoxic pentacyclic triterpenoids from *Combretum sundaicum* and *Lantana camara* as inhibitors of Bcl-xL/BakBH3 domain peptide interaction. J Nat Prod. 2009;72(7):1314-1320.

DOI: 10.1021/np900192r.

 Viau CM, Moura DJ, Facundo VA, Saffi J. The natural triterpene 3β,6β,16βtrihydroxy-lup-20(29)-ene obtained from the flowers of Combretum leprosum induces apoptosis in MCF-7 breast cancer cells. BMC Complement Altern Med. 2014;14:280.

DOI: 10.1186/1472-6882-14-280.

- 59. Pettit GR, Cragg GM, Singh SB. Antineoplastic agents, 122. constituents of *Combretum caffrum*. J Nat Prod. 1987;50(3):386-391.
- 60. Fyhrquist P, Salih EYA, Helenius S, Laakso I, Julkunen-Tiitto R. HPLC-DAD and UHPLC/QTOF-MS analysis of polyphenols in extracts of the African species *Combretum padoides*, C. zeyheri and C. psidioides related to their antimycobacterial activity. Antibiotics (Basel). 2020;9(8):459.

DOI: 10.3390/antibiotics9080459.

61. Anokwuru CP, Chen W, van Vuuren S, Combrinck S, Viljoen AM. Bioautographyguided HPTLC-MS as a rapid hyphenated technique for the identification of antimicrobial compounds from selected South African *Combretaceae species*. Phytochem Anal. 2022;33(8):1177-1189. DOI: 10.1002/pca.3167.

 Karatoprak GŞ, Küpeli Akkol E, Genç Y, Bardakci H, Yücel Ç, Sobarzo-Sánchez E. Combretastatins: An overview of structure, probable mechanisms of action and potential applications. Molecules. 2020; 25(11):2560.

DOI: 10.3390/molecules25112560.

- 63. Lawrence NJ, Ghani FA, Hepworth LA, Hadfield JA, McGown AT, Pritchard RG. The synthesis of (E) and (Z)combretastatins A-4 and a phenanthrene from *Combretum caffrum*. Synthesis 1999;9:1656-1660.
- 64. Vitale I, Antoccia A, Cenciarelli C, Crateri P, Meschini S, Arancia G et al. Combretastatin CA-4 and combretastatin derivative induce mitotic catastrophe dependent on spindle checkpoint and caspase-3 activation in non-small cell lung cancer cells. Apoptosis. 2007;12(1):155-166.

DOI: 10.1007/s10495-006-0491-0.

- Barnes NG, Parker AW, Ahmed Mal Ullah AA, Ragazzon PA, Hadfield JA. A 2-step synthesis of *Combretastatin* A-4 and derivatives as potent tubulin assembly inhibitors. Bioorg Med Chem. 2020;28(19):115684.
 DOI: 10.1016/j.bmc.2020.115684
- Pettit GR, Singh SB, Hamel E, Lin CM, Alberts DS, Garia-Kendall D. Isolation and structure of the strong cell growth and tubulin inhibitor combretastatin A4. Experientia. 1989;45:205-211. DOI: 10.1007/BF01954881
- 67. McGown AT, Fox BW. Differential cytotoxicity of *Combretastatins* A1 and A4 in two daunorubicin-resistant P388 cell lines. Cancer Chemother Pharmacol. 1990;26:79-81.

Available:https://doi.org/10.1007/BF02940 301

68. West CM, Price P. Combretastatin A4 phosphate. Anticancer Drugs. 2004;15(3): 179-187.
DOI:10.1097/00001813-200403000-00001.

69. Liu P, Qin Y, Wu L, Yang S, Li N, Wang H, et al. A phase I clinical trial assessing the safety and tolerability of combretastatin A4 phosphate injections. Anticancer Drugs. 2014;25(4):462-471.

70. Grisham R, Ky B, Tewari KS, Chaplin DJ, Walker J. Clinical trial experience with CA4P anticancer therapy: Focus on efficacy, cardiovascular adverse events, and hypertension management. Gynecol Oncol Res Pract. 2018;5:1.

DOI: 10.1186/s40661-017-0058-5.

71. Gossan DPA, Magid AA, Yao-Kouassi PA, Josse J, Gangloff SC, Morjani H, et al. Antibacterial and cytotoxic triterpenoids from the roots of *Combretum racemosum*. Fitoterapia. 2016;110:89-95.

> Available:https://doi.org/10.1016/j.fitote.20 16.03.002

- 72. Moosophon P, Kanokmedhakul S, Kanokmedhakul K, Buayairaksa M, Noichan J, Poopasit K. Antiplasmodial and cytotoxic flavans and diarylpropanes from the stems of *Combretum griffithii*. J Nat Prod. 2013;76(7):1298-1302. DOI: 10.1021/np400266h.
- Banskota AH, Tezuka Y, Phung LK, Tran KQ, Saiki I, Miwa Y et al. Cytotoxic cycloartane-type triterpenes from *Combretum quadrangulare*. Bioorg Med Chem Lett. 1998;8(24):3519-3524.
 DOI: 10.1016/s0960-894x(98)00644-1.
- 74. Vo TS, Nguyen HH, Nguyen TP, Tran TM, Bui XH, Dinh MH et al. Cycloartanes from leaves of *Combretum quadrangulare* growing in Vietnam. Nat Prod Res. 2022;1-8.

DOI: 10.1080/14786419.2022.2045489.

Banskota AH, Tezuka Y, Tran KQ, Tanaka 75. Saiki Kadota S. Methyl Κ, Ι, quadrangularates A-D related and triterpenes from Combretum guadrangulare. Chem Pharm Bull (Tokyo). 2000;48(4):496-504.

DOI: 10.1248/cpb.48.496.

 Simon G, Dewelle J, Nacoulma O, Guissou P, Kiss R, Daloze D, et al. Cytotoxic pentacyclic triterpenes from *Combretum nigricans*. Fitoterapia. 2003;74(4):339-344.

DOI: 10.1016/s0367-326x(03)00046-7.

- Wu XP, Han CR, Chen GY, Yuan Y, Xie JY. Cytotoxic pentacyclic triterpenoids from *Combretum oliviforme*. Nat Prod Commun. 2010;5(7):1027-1030.
- Wende M, Sithole S, Chi GF, Stevens MY, Mukanganyama S. The effects of combining cancer drugs with compounds isolated from *Combretum zeyheri* Sond. and *Combretum platypetalum* Welw. ex M.A. Lawson (Combretaceae) on the viability of Jurkat T cells and HL-60 cells. Biomed Res Int; 2021.

DOI: 10.1155/2021/6049728.

79. Bisoli E, Freire TV, Yoshida NC, Garcez WS, Queiróz LMM, Matos MFC, et al. Cytotoxic phenanthrene, dihydrophenanthrene, and dihydrostilbene derivatives and other aromatic compounds from *Combretum laxum*. Molecules 2020; 25(14):3154.

DOI: 10.3390/molecules25143154.

- Williams RB, Norman VL, Goering MG, O'Neil-Johnson M, Eldridge GR, Starks CM. Acetylated dammarane-type bisdesmosides from *Combretum inflatum*. J Nat Prod. 2013;76(9):1592-1597.
 DOI: 10.1021/np4002652.
- Mirowski PDS, Ojeda M, Kollet LG, Freire TV, Pott A, Garcez WS, et al. Selective tumor cell growth inhibition by lignans and a seco-triterpenoid from *Combretum mellifluum*. Nat Prod Res. 2022;10:1-8.

DOI: 10.1080/14786419.2021.2024823

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