Protective effect of lawsone on L-Arginine induced acute pancreatitis in rats

Sandeep Biradar* & B Veeresh

Department of Pharmacology, G. Pulla Reddy College of Pharmacy, Mehdipatnam, Hyderabad 500 028, India

Received 20 June 2012; revised 11 December 2012

The efficacy of lawsone against L-arginine induced acute pancreatitis was determined at 24 h by determination of serum levels of amylase, lipase and proinflammatory cytokines [tumor necrosis factor (TNF)- α , C-reactive proteins and interleukin (IL)], pancreatic myeloperoxidase (MPO) activity, lipid peroxidation (thiobarbituric acid reactive substances (TBARS)], nitrate/nitrite levels, and the wet weight/body weight ratio. Lawsone and methylprednisolone treatments significantly attenuated the L-arginine- induced increases in pancreatic wet weight/body weight ratio, and decreased the serum levels of amylase and lipase, and TNF- α and IL-6 and significantly lowered pancreatic levels of MPO, TBARS, and nitrate/nitrite. The histoimmunological findings further proved the amelioration of pancreatic injury by lawsone and further proved anti-inflammatory and antioxidant agent property of lawsone.

Keywords: Acute pancreatitis, Anti-inflammatory, Cytokines, L-arginine, Lawsone

Inflammation of pancreatic gland called pancreatitis (AP), may leads to sever complication and high mortality without treatment. The pathogenesis is not fully understood, however the leukocyte activation, microcirculatory disturbances and oxidative stress are the major constituents of AP. This is characterized by activation widespread inflammatory of cell infiltration, leukocyte and digestive proteases. Further, by releasing reactive oxygen, nitrogen species and various kinds of inflammatory mediators. Several factors are responsible for the AP, like hereditary alcohol. gallstones, pancreatitis, hypercalcemia, hyperlipidemia, malnutrition, abdominal trauma, penetrating ulcers, malignancy, drugs like steroids, sulfonamides, furosemide, thiazides, infections like mumps, coxsackie virus, mycoplasma pneumoniae, ascaris, Clonorchis, and structural abnormalities like choledochocele and pancreas divisum. Repeated attacks of acute pancreatitis have the potential to develop into chronic pancreatitis or pancreatic cancer characterized by fibrosis and loss of acinar cell function^{1,2}. No specific treatment is available to treat AP. Many therapies and medical management is aimed to control the sign and symptoms of AP, using steroids, analgesics and antiinflammatory agents. The use of the synthetic and semi-synthetic treatment has various kinds of drawbacks like photosensitivity skin reactions, intolerance and addiction. Apart from this, these compounds are very expensive and not reliable. Hence, there is need to explore potential antioxidant and anti-inflammatory agents available from natural sources, which are cost-effective and have several advantages than the synthetic and semisynthetic compounds. Antioxidant, anti-inflammatory and anticarcinogenic activities of lawsone have been reported^{3,4}. The lawsone has haematotoxic properties, leading to stimulation of cell proliferation; representing a sufficient explanation for a weak induction of 'late micronuclei'. Hence in this study the non-toxic dose of lawsone (100 and 200 mg/kg) have been used to evaluate the potential effect to ameliorate pancreatic injury induced by L-arginine.

Materials and Methods

Animals—Male Wistar rats (30) weighing 180-200 g obtained from Mahaveer Enterprises, Hyderabad were maintained at a constant room temperature $(23\pm2 \ ^{\circ}C)$ with 12:12 h light-dark cycles and free access to water and standard laboratory chow. The rats were randomly divided into 6 groups of 8 in each and experiments were performed after 12 h of fasting. All the experimental procedures were carried out in accordance with Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA) guidelines. The study was

^{*}Correspondent author Cell: +91 8978517068 E-mail: sandeepbiradar84@gmail.com

reviewed and approved by the Institutional Animal Ethics Committee (320/CPCSEA dated 03-01-2001), G.Pulla Reddy College of Pharmacy, Hyderabad, India.

Chemicals—L-arginine (Sigma Aldrich Co Pvt Ltd, USA), lawsone (Sigma Aldrich Co Pvt Ltd, Japan), hexadecyltrimethylammonium bromide (HETAB) (Sigma Aldrich Co Pvt Ltd, Switzerland), o-dianisidinedihydrochloride, thiobarbituric acid (TBA) (Sigma Aldrich Co Pvt Ltd, Germany), Griess reagent (Sigma Aldrich Co Pvt Ltd, Germany) and vanadium trichloride (Sigma Aldrich Co Pvt Ltd, USA) were procured from Sigma Aldrich Chemical Co. All other chemicals and reagents were of highest commercial grade available locally.

L-arginine, powder: prepared as a solution by dissolving in 0.9% saline to a final concentration of 500 mg/mL and the pH was adjusted to 7 with 5 N HCl.

Lawsone was prepared as a solution by dissolving in 3% tween 80 and 0.9% saline to a final concentration of 100 and 200 mg/mL and the pH was adjusted to 7 with 0.1 N NaOH.

L-arginine-induced pancreatitis model—Acute pancreatitis was induced in five groups of rats by two intraperitoneal (ip) injections of L-arginine (2.5 g/kg, 1 h apart). One hour following the last injection of L-arginine, the rats were treated orally as follows: Gr. 1 received the vehicle (3% Tween 80) of lawsone (vehicle control); Grs 2 and 3 were treated with lawsone (100 and 200 mg/kg, respectively). Gr. 4 acted positive control and received as methylprednisolone (30 mg/kg), all in a volume of 10 mL/kg and Gr. 5, received saline (0.9%, NaCl, ip) in place of L-arginine and served as a normal control. After 24 h of the last injection of L-arginine or saline, a midline laparotomy was performedin rats under ether anesthesia and blood samples were collected from the inferior vena cava, the rats were then exsanguinated, the whole pancreas was quickly removed and stored at -70 °C until use. The pancreatic weight/body weight ratio was evaluated as an estimate of the degree of pancreatic edema (mg/g).

Macroscopic evaluation

Pancreas weight/body weight ratio—The pancreas was removed immediately after the blood collection, trimmed free of fat and weighed. The pancreatic weight/body weight ratio (mg/g) was calculated for each animal, to estimate the level of pancreatic edema⁵.

Serum analysis—For serum analysis, blood samples were centrifuged at 3000 g at 4 °C for 10 min. The serum amylase and lipase were

determined by routine colorimetric methods using the commercial kits for amylase (Rapid diagnostics), lipase (Accurex diagnostics), C-reactive protein and interleukin- α and expressed as U/dl⁵.

Biochemical estimations—Pancreatic total protein content⁶ was determined. Myeloperoxidase activity⁷, nitrate/nitrite level^{8,9}, TBARS level¹⁰, catalase activity^{11,12}, SOD level¹³ and reduced GSH level¹⁴ were measured.

Histopathological evaluation-Pancreas was removed immediately and part of it was fixed in 10% neutral buffered formalin and embedded in paraffin by standard methods. Paraffin sections of 5 µm thicknesses were cut and stained with haematoxylin and eosin, assessed under dark field microscope and examined blind by a morphologist for grading histopathological changes. Pancreatic damage was assessed and scored by grading acinar cell degeneration, interstitial inflammation, edema, and haemorrhage as described by schmidt's standards^{15,16} with modification as follows: Grading for edema was scaled as 0: absent or rare; 1: edema in the interlobular space; 2: edema in the intralobular space; 3: isolated island shape of pancreatic acinus. Inflammation was scored as 0: absent; 1: mild; 2: moderate: 3: severe. Acinar cell necrosis was scored as 0: absent; 1: mild; 2: moderate; 3: severe. Parenchyma haemorrhage was scored as 0: absent; 1: mild; 2: moderate; 3: severe. The maximum score for acinar cell damage was 12.

Statistical analysis—Statistical analysis was performed by one way ANOVA followed by Newman Keuls as post-hoc test using Graph pad Prism 5. Values were presented as mean \pm SE. The difference was considered to be statistically significant when P < 0.05.

Results

Serum biochemical parameters and pancreatic edema—Induction of pancreatitis resulted in significant raise in the serum amylase, lipase and pancreatic edema. Treatment with lawsone (100 and 200 mg/kg) dose dependently decreased the serum amylase, lipase and pancreatic edema (Table 1).

Pancreatic MPO and total protein—Induction of pancreatitis resulted in significantly increased the pancreatic MPO and decreased the pancreatic total protein levels. Treatment with lawsone (100 and 200 mg/kg) dose dependently reversed the change in pancreatic MPO and total protein levels (Table 1).

Pancreatic, lung, liver and kidney MDA, nitrate/nitrite, GSH and antioxidant enzymes catalase

and SOD—Induction of pancreatitis resulted in a significant raise in MDA, nitrate/nitrite, catalase and SOD and decline in GSH levels. Treatment with lawsone (100 and 200 mg/kg) dose dependently reversed the change in MDA, nitrate/nitrite, catalase, SOD and GSH levels (Table 1).

Assessment of interleukins and C-reactive protein—Induction of pancreatitis resulted in a significant raise in interleukins, TNF- α and C- reactive protein. Treatment with lawsone (100 and 200 mg/kg) dose dependently decreased the interleukins and C- reactive protein (Table 2).

Pancreatic histology-Histological examination of normal control group (saline treated) showed normal architecture and absence of edema, neutrophil infiltration, hemorrhage and necrosis (Fig. 1). Whereas, pancreatic sections of disease control group showed extensive tissue damage characterized by acinar cell degeneration, necrosis, edema. mononuclear cell infiltration. hemorrhage and thus received significantly higher scores. Treatment with lawsone (100 and 200 mg/kg) and methyl prednisolone (30 mg/kg) ameliorated the inflammation, edema and more significantly acinar cell degeneration and necrosis and protected the pancreas from L-arginine induced damage. Treatment with lawsone dose dependently decreased the total pathological scores compared to disease control group.

Table1—Effect of lawsone on pancreas weight, total body weight, serum amylase, serum lipase, total nitrate, total protein, MDA, MPO and SOD after L-arginine induced acute pancreatitis [Values are mean ± SE from 6 animals in each group]								
Parameter/Groups	NC	DC	STD	LW 100	LW 200			
Pancreas weight	15.36±870.3	19.04±1015*	18.83±911.3α	$46.33 \pm 843.3 \alpha$	21.37±788.3α			
Total body wt	4.889±187.5	$3.578 \pm 192*$	3.60 ± 191.2	5.57 ± 188.7	3.68 ± 189			
Serum Amylase	2000±85.63	7667±349*	3317±110.8α	2733±140.6α	3167±187.4α			
Serum Lipase	191.7±4.014	566.7±30.84*	346.7±39.21α	$236.7 \pm 14.06\alpha$	260±7.303α			
Total Nitrate	1.372±11.87	1.462±16.07*	$0.39 \pm 7.06 \alpha$	$1.31\pm10.13\alpha$	$0.77\pm8.5\alpha$			
Total Protein	0.032 ± 0.73	0.037±0.355*	$0.067 \pm 0.91 \alpha$	$0.02\pm0.73\alpha$	$0.05\pm0.82\alpha$			
Kidney GSH	0.011 ± 0.47	0.04± 0.284 *	$0.014 \pm 0.71 \alpha$	$0.017 \pm 0.51\beta$	$0.049 \pm 0.64 \alpha$			
Liver GSH	0.010 ± 0.48	0.04 ±0.284 *	$0.03 \pm 0.67 \alpha$	$0.03 \pm 0.48 \beta$	$0.04\pm0.59\alpha$			
Lung GSH	0.020 ± 0.50	0.04± 0.284 *	$0.03\pm0.67\alpha$	$0.02 \pm 0.51\gamma$	$0.04\pm0.60~\beta$			
Pancreas GSH	0.011 ± 0.48	0.04± 0.284 *	$0.03\pm0.67\alpha$	$0.02 \pm 0.52 \ \beta$	$0.04\pm0.57\alpha$			
Kidney MDA	0.92 ± 13.91	1.04± 17.09 *	$0.52 \pm 9.61\beta$	$0.68 \pm 12.31 \alpha$	$0.39 \pm 10.72 \alpha$			
Liver MDA	0.88 ± 14.36	1.04 ±17.09 *	$0.62 \pm 10.96 \gamma$	$0.85 \pm 13.97 \alpha$	$0.62 \pm 12.56c$			
Lung MDA	0.66 ± 14.95	1.04 ± 17.09	$1.04 \pm 9.82\beta$	$1.70 \pm 13.18 \alpha$	$0.66 \pm 11.5\alpha$			
Pancrease MDA	1.62±0.6615	1.04±0.4267*	2.39±0.9756α	1.70±0.69 β	1.21±0.49 β			
Pancreas MPO	4.75 ± 2.1	$32.8\pm4.7*$	$6.2 \pm 2.2 \alpha$	$17.2 \pm 3.7 \alpha$	$11.2 \pm 3.7 \alpha$			
Lung MPO	8.3 ± 2.4	$43.7 \pm 7.1*$	$11.1 \pm 3.0\alpha$	$28.4\pm3.3\alpha$	13.4 ± 3.3 g			
Liver MPO	7.75 ± 2.1	$33.8 \pm 4.7*$	$9.2\pm2.2\alpha$	$22.2\pm3.4\alpha$	$14.2 \pm 3.8\alpha$			
KIdney MPO	5.3 ± 2.4	$39.7 \pm 7.1*$	$8.2\pm3.0\alpha$	$26.4\pm3.5\alpha$	$13.4 \pm 3.6 \alpha$			
Pancreas Catlaze	0.33±0.13	0.41±0.1707*	0.19±0.07	0.23±0.0934β	0.23±0.0948α			
kidney SOD	2.33 ± 0.95	1.50 ± 0.6146	2.61±1.06α	1.89 ± 0.774	1.891±0.771β			
Liver SOD	0.83±0.34	1.01±0.4153	$1.97\pm0.80\alpha$	1.527±0.62β	1.78±0.7266β			
Lung SOD	1.04 ± 0.42	1.03±0.42*	$3.44{\pm}1.40\alpha$	2.06±0.84	3.83±1.563α			
Pancrease SOD	2.09 ± 0.85	1.51±0.6191*	1.49±0.61 α	1.32±0.54	1.81±0.74			
< 0.0001 when compared	with normal control, ap	< 0.0001, βp< 0.001,	γp< 0.01 when comp	ared with disease con	trol group			

Table 2—Effect of lawsone on Interleukins values, C - reactive protein and TNF- α [Values are mean ± SEM from 6 animals in each group]							
Serum parameters	NC	DC	LW1	LW2			
Interlukin-6	29.3±1.569	90.48±1.689*	62.42±1.358γ	33.57±1.182γ			
TNF-α	19.33±1.541	26.32±3.036	21.58±0.9711	18.57±1.316			
C-Reactive protein	415.3±7.762	16403±119*	8611±101.7γ	546.8±9.874 γ			
N C- Normal Control	D C- Disease Control L V	/ 100-Lawsone 100 mg/kg LV	W 200- Lawsone 200 mg/kg 3	*P < 0.0001 when compared			

N.C- Normal Control, D.C- Disease Control, LW 100- Lawsone 100 mg/kg, LW 200- Lawsone 200 mg/kg. *P < 0.0001 when compared with normal control, $\gamma P < 0.01$ when compared with disease control group.

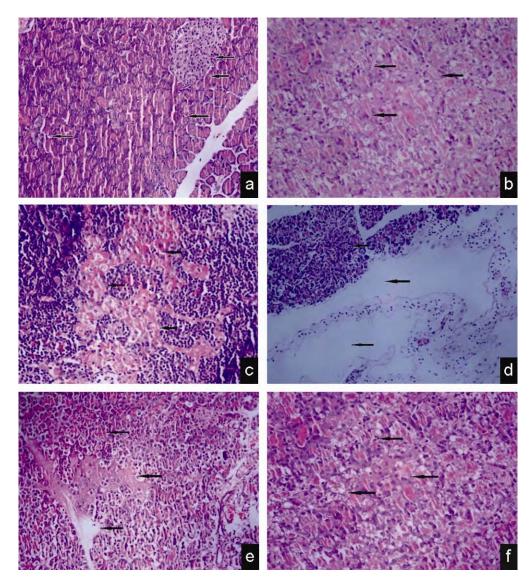


Fig. 1—Effect of lawsone on pancreatic histopathological changes after L-arginine induced acute pancreatitis [(a) normal control, (b) standard control, (c) disease control 24 h hemorrhage, (d) disease control 24 h edema and necrosis, (e) lawsone 100 mg/ kg, ip, 24 h, (f) lawsone 200 mg/ kg, ip, 24 h] (H&E \times 200)

Discussion

The present study demonstrated that treatment with lawsone (200 mg/kg) efficiently reduced the severity of L-arginine induced acute pancreatitis in rats. In consistent with previous reports^{2,17,18}, in the present study administration of L-arginine significantly developed the acute pancreatitis characterized by raised levels of serum amylase, lipase and acinar cell necrosis.

Serum amylase and lipase levels are the important diagnostic markers for acute pancreatitis. They usually rise within 4–8 h of the initial attack, peaks at 24 h and returns to normal over the 72 $h^{5,6,17}$. Similarly, in accordance with previous reports, in the present study induction of pancreatitis significantly

increased the serum amylase and lipase levels at 24 h. Treatment with lawsone decreased the serum amylase and lipase levels, indicating protective effect of lawsone at early stage of the disease progression.

In consistent with previous reports^{5,6,17}, in the present study induction of pancreatitis significantly increased the pancreatic MPO, MDA, nitrite, catalase and SOD and decreased the GSH levels. MPO, a marker of neutrophil infiltration is an enzyme found in neutrophils and its activity is linearly related to infiltration of neutrophils^{19,20}. In agreement with previous reports^{5,21}, induction of pancreatitis with L-Arginine increased the pancreatic MPO levels. Inhibition of the neutrophil infiltration can attenuate

the pancreatic injury²². Treatment with lawsone significantly decreased the pancreatic MPO levels probably due to its anti-inflammatory action.

MDA, a marker of lipid peroxidation was elevated in L-arginine treated rats. Lipid peroxidation is a process mediated by free radicals, which results in impairment of the membrane functional and structural integrity^{6,17,23} resulting in oxidative deterioration of polyunsaturated fatty acids of cell membrane. It could be attributed to the accumulation of free radicals proposed to be generated by L-arginine. The change in levels of catalase and SOD remains controversial. Czako and Takacs²³ reported the fall in these enzyme levels at 24 h. However, Szabolcs et al.¹⁷ reported the raised levels of these enzymes. In consistent with Robbins², in the present study significant increase in SOD and catalase level was observed. It indicates that oxidative stress caused by L-arginine may up-regulate the activity of antioxidant enzymes to facilitate rapid removal of accumulated reactive oxygen and nitrogen species¹⁷. It is well known that GSH is found to be decreased in L-arginine treated rats indicating enhanced oxidative stress as the disease progresses¹⁷.

The role of NO in the initiation and progression of acute pancreatitis remains controversial²⁴. Some studies²⁵⁻²⁸ reported that NO increase the pancreatic blood flow and/or secretion in response to endothelium derived NO and ameliorates the pancreatic dysfunction, others²⁹⁻³¹ suggested that NO aggravates pancreatic oxidative stress and damage. In agreement with previous reports³¹, in the present study significant increase in NO and pancreatic edema was observed in L-arginine received rats. Takacs *et al.*³¹ demonstrated that, administration of excess L-arginine could induce iNOS activity and increase the NO levels in pancreas. The raised levels of NO can increase vascular/micro capillary permeability and may contribute to the pancreatic edema and acinar cell damage.

Treatment with lawsone significantly restored the pancreatic MDA, nitrite, edema, catalase, SOD and GSH in L-arginine received rats. Passaglia³² stated that acinar cells are the protein factory of the body. In acute pancreatitis, catabolism of proteins could increase up to 80%. Consequently, a sharp decline in protein content was observed in pancreas. In consistent with Sidhu *et al.*⁶, pancreatic total protein content, a marker of the tissue damage was found to decrease in L-arginine received rats. Treatment with lawsone significantly increased the total protein content.

It is well known that the extent of pancreatic tissue damage in acute pancreatitis correlates with the levels of inflammatory mediators and free radicals. In agreement with previous reports^{5,6,21}, in the present study, histopathological assessments revealed that, induction of pancreatitis resulted in pancreatic damage characterized by acinar cell necrosis, mono nuclear cell infiltration, edema and haemorrhage. Treatment with lawsone protected the pancreas from L-arginine induced injury. In conclusion, the present study suggests that treatment with lawsone significantly ameliorated the severity of L-arginine induced pancreatitis by reducing the neutrophil infiltration and oxidative stress markers and this effect may be due to antioxidant and anti-inflammatory properties of the lawsone.

Disclaimer

The views and opinions expressed in this article are those of the authors, and they do not reflect in any way those of the institutions to which they are affiliated with.

References

- 1 Robbins K, *Basic pathology (Elsevier Publications, Chicago)* 1997, 902.
- 2 Robbins K, *Basic pathology*, (*Elsevier Publications*, Chicago) 2002, 675.
- 3 Sauriasari R, Wang DH, Takemura Y, Tsutsui K, Masuoka N, Sano K, Horita M, Wang BL & Ogino K, Cytotoxicity of lawsone and cytoprotective activity of antioxidants in catalase mutant *Escherichia coli*, *Toxicology*, 235(1-2) 2007, 103,
- 4 Mikhaeil BR, Badria FA, Maatooq GT & Amer MMA, Antioxidant and immunomodulatory constituents of henna leaves, *Z. Naturforsch*, 59 c (2004) 468.
- 5 Melo CM, Carvalho KMMB, Neves JCDS, Morais TC, Rao VS, Santos FA, Brito GADC & Chaves MH, α,β-amyrin, a natural triterpenoid ameliorates L-arginine induced acute pancreatitis in rats, *World J Gastroenterol*, 16 (34), (2010) 4272.
- 6 Sidhu S, Pandhi P, Malhotra S, Vaiphei K & Khanduja KL, Melatonin treatment is beneficial in pancreatic repair process after experimental acute pancreatitis, *Eur J Pharmacol* 628 (2010) 282.
- 7 Bradley PP, Priebat DA, Christensen RD & Rothstein G, Measurement of cutaneous inflammation: estimation of neutrophil content with an enzyme marker, J Invest Dermatol, 78 (1982) 206.
- 8 Green LC, Wagner DA, Glogowski J, Skipper PL, Wishnok JS &Tannenbaum SR, Analysis of nitrate and nitrite in biological fluids, *Anal Biochem*, 126 (1982) 131.
- 9 Gamal el-din AM, Mostafa AM, Al-Shabanah OA, Al-Bekairi AM & Nagi MN, Protective effect of arabic gum against acetaminophen-induced hepatotoxicity in mice, *Pharmacol Res*, 48 (2003) 631.
- 10 Ohkawa H, Ohishi N & Yagi K, Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction, *Anal Biochem*, 95 (1979) 351.

- 11 Aebi H, Catalase, *in Methods in enzymology*, edited by L Packer (Academic Press, Orlando) Vol 105, 1984, 121.
- 12 Kakkar R, Mantha SV, Radhi J, Prasad K & Kalra J, Increased oxidative stress in rat liver and pancreas during progression of streptozotocin-induced diabetes, *Clin Sci*, 94 (1998) 623.
- 13 Misra HP & Fridovich I, The role of superoxide anion in the autoxidation of epinephrine and a simple assay for superoxide dismutase, *J Biol Chem*, 247 (1972) 3170.
- 14 Sedlak J & Lindsasy RH, Estimation of total, protein-bound, and nonprotein sulfhydril groups in tissue with Ellman's reagent, *Anal Biochem*, 25 (1968) 192.
- 15 Schmidt J, Rattner DW, Lewandrowski K, Compton CC, Mandavilli U, Knoefel WT & Warshaw AL, A better model of acute pancreatitis for evaluating therapy, *Ann Surg*, 215 (1992) 44.
- 16 Eşrefoglu M, Gül M, Ates B, Batçioglu K & Selimoglu MA, Antioxidative effect of melatonin, ascorbic acid and Nacetylcysteine on caerulein-induced pancreatitis and associated liver injury in rats, *World J. Gastroenterol*, 12 (2006) 259.
- 17 Szabolcs A, Reiter RJ, Letoha T, Hegyi P, Papai G, Varga I, Jarmay K, Kaszaki J, Sari R, Rakonczay Z Jr, Lonovics J & Takacs T, Effect of melatonin on the severity of L-arginineinduced experimental acute pancreatitis in rats, *World J Gastroenterol*, 12 (2006) 251.
- 18 Hegyi P, Rakonczay Z Jr, Sari R, Góg C, Lonovics J, Takacs T &Czako L, L-arginine induced experimental pancreatitis, *World J. Gastroenterol* 10 (14) (2004) 2003.
- 19 Poch B, Gansauge F, Rau B, Wittel U, Gansauge S, Nüssler AK, Schoenberg M & Beger HG, The role of polymorphonuclear leukocytes and oxygen-derived free radicals in experimental acute pancreatitis: mediators of local destruction and activators of inflammation, *FEBS Lett*, 461 (1999) 268.
- 20 Song AM, Bhagat L, Singh VP, Van Acker GG, Steer ML & Saluja AK, Inhibition of cyclooxygenase-2 ameliorates the severity of pancreatitis and associated lung injury, *Am J Physiol Gastrointest Liver Physiol*, 283 (2002) 1166.

- 21 Abdin AA, El-Hamid MA, El-Seoud SH & Balaha MF, Effect of pentoxifylline and/or alpha lipoic acid on experimentally induced acute pancreatitis, *Eur J Pharmacol*, 643 (2010) 289-296.
- 22 Shi C, Zhao X, Wang X, Zhao L & Andersson R, Potential effects of PKC or protease inhibitors on acute pancreatitis induced tissue injury in rats, *Vascul Pharmacol*, 46 (2007) 406.
- 23 Czako L & Takacs T, Involvement of oxygen derived free radicals in L-arginine induced acute pancreatitis, *Dig Dis Sci*, 43 (8)(1998) 1770.
- 24 Sweiry JH & Mann GE, Role of oxidative stress in the pathogenesis of acute pancreatitis, *Scand J Gastroenterol*, 31 (1996) 10.
- 25 Gukovskaya A & Pandol S, Nitric oxide production regulates cGMP formation and calcium influx in pancreatic acinar cells, *Am J Physiol*, 266 (1994) 350.
- 26 Holst JL, Rasmussen TN & Scmidt P, Role of nitric oxide in neutrally induced pancreatic exocrine secretion in pigs, *Am J Physiol*, 266 (1994) 206.
- 27 Satoh A, Shimosegawa T, Abe T, Kikuchi Y, Abe R, Koizumi M & Toyota T, Role of nitric oxide in the pancreatic blood flow response to caerulein, *Pancreas*, 9 (1994) 574.
- 28 Patel AG, Toyama MT, Nguyen TN, Cohen GA, Ignarro LJ, Reber HA & Ashley SW, Role of nitric oxide in the relationship of pancreatic blood flow and exocrine secretion in cats, *Gastroenterology*, 108 (1995) 1215.
- 29 Tani S, Itoh H & Okabayashi Y, New model of acute necrotizing pancreatitis induced by excessive doses of arginine in rats, *Dig Dis Sci* 35(3) (1990) 367.
- 30 Dabrowski A & Gabryelewicz A, Nitric oxide contributes to multi-organ oxidative stress in acute pancreatitis, *Scand J Gastroenterol*, 29 (1994) 943.
- 31 Takacs T, Czako L, Morschl E, Laszlo F, Tiszlavicz L, Rakonczay Z Jr & Lonovics J, The role of nitric oxide in edema formation in L-arginine-induced acute pancreatitis, *Pancreas*, 25 (2002) 277.
- 32 Passaglia C, Nutritional problems in acute pancreatitis, *Recent Prog Med*, 98 (2007) 335.