

DETECTION OF OPPORTUNISTIC DNA VIRAL INFECTIONS BY MULTIPLEX PCR AMONG HIV INFECTED INDIVIDUALS RECEIVING CARE AT A TERTIARY CARE HOSPITAL IN SOUTH INDIA

J Sachithanandham, M Ramamurthy, *R Kannangai, HD Daniel, OC Abraham, P Rupali, SA Pulimood, AM Abraham, G Sridharan

Abstract

Purpose: Opportunistic viral infections cause increased morbidity and mortality among human immunodeficiency virus (HIV) infected individuals, especially those who are not on antiretroviral treatment. Early diagnosis of these opportunistic viruses will be able to reduce the risk of disease progression with appropriate intervention. **Materials and Methods:** Multiplex PCR was attempted to detect the opportunistic herpes viruses (HSV-1, HSV-2, VZV, EBV, and CMV), adenovirus and polyoma viruses (JC and BK) in three cocktails of PCR reactions. Subsequently, all the viruses detected were quantitated by testing using monoplex real time PCR. Whole blood samples collected between 2006 and 2007 from 68 treatment naïve HIV-1 infected and 30 normal healthy individuals were tested for these eight viruses. Among the 68 HIV-1 infected individuals 35 had CD4+ T cell count less than or equal to 200 while the other 33 had greater than 200 CD4+ T cells. **Results:** Among the 68 HIV-1 infected individuals, 49 (72%) were positive for EBV, 5 (7%) samples were positive for CMV. All the five CMV positive individuals had CD4+ T cell count of less than or equal to 200 cells/ μ L. The mean EBV load among the individuals with a CD4+ T cells of less than or equal to 200 cells/ μ L was 3.88 \log_{10} while among those with greater than 200 CD4+ T cells it was 3.75 \log_{10} . The mean CMV load was 6.98 \log_{10} . Three samples were positive for both CMV & EBV. None of the samples was positive for HSV-1, HSV-2, VZV, Adenovirus, JC and BK viruses. **Conclusions:** In our study, multiplex PCR based detection system was found useful in detecting opportunistic viruses in HIV infected individuals. Though EBV is the most prevalent opportunistic viral infection among HIV infected individuals, there was no significant association between EBV load, CD4+ T cell counts and HIV-1 virus load. CMV was seen in HIV infected individuals with low CD4+ T cell counts (less than 200 cells/ μ L).

Key words: Human immunodeficiency virus, India, multiplex polymerase chain reaction, opportunistic viruses

Introduction

In HIV infected individuals, opportunistic viral infections are the major cause of morbidity and mortality. These agents cause infections which could be asymptomatic or mildly symptomatic in immunocompetent individuals, and it is often self-limiting. However, in immunosuppressed individuals and individuals with malignancy, infection with these agents leads to severe life-threatening diseases.^[1,2]

The primary target of HIV during infection is CD4+ cells that lead to reduction in CD4+ T cells and thereby cause immunosuppression. During HIV disease progression infection, there will be reduction of CD4+T cells and the infected individuals could succumb to AIDS related opportunistic viral infections when CD4 cells are less than

200cells/ μ L.^[3] The important opportunistic viruses that can cause infection in HIV infected individuals are herpes viruses like HSV, VZV, EBV and CMV. Herpes virus infections are seen in HIV infected individuals and can manifest as meningitis, encephalitis, oral hairy cell leukoplakia (OHL), lymphoproliferative disorders, retinitis.^[4,5] One another important herpes virus that can cause Kaposi's sarcoma associated lesions in HIV infected individuals is the human herpes virus-8 (HHV-8).^[6]

Polyomaviruses like the JC virus causes progressive multifocal leucoencephalopathy in HIV infected individuals.^[7] Infections associated with adenovirus in HIV infected individuals manifest as lethal pneumonia, diarrhoea and encephalitis.^[8] In HIV infected individuals, the detection of these opportunistic viruses and specific treatment are very important to effectively prevent their deleterious effects.

There were several techniques to detect these opportunistic viruses, such as cell culture for isolation, Enzyme Linked Immunosorbent Assay (ELISA) for antigen or antibody detection, and Polymerase Chain Reaction (PCR) for nucleic acid detection.^[9,10] Cell culture isolation has limited scope because some viruses cannot easily be grown in culture (JC and BK) and is also time consuming. ELISA, though not expensive, has some drawbacks like

*Corresponding author (email: <rajeshkannangai@hotmail.com>)
Department of Clinical Virology (JS, MR, RK, HDD, AMA, GS),
Department of Internal Medicine (OCA, PR), Department of
Dermatology (SAP), Christian Medical College, Vellore - 632004,
India
Received: 28-12-2008
Accepted: 07-05-2009

pre-exposure to viruses like CMV; EBV reduces the specificity for the episode of illness. The PCR is very useful in screening for these opportunistic viruses and a multiplex assay has an additional advantage of screening for these opportunistic viruses since it involves the simultaneous amplification of one or more targets. It also enables a wide range of samples to be tested, like blood, CSF, urine.^[10] Many recent studies have shown the importance of using whole blood rather than fractions for testing for viral infections, especially in the case of CMV.^[11] Early detection is very important in treating opportunistic infections and there are many drugs to improve the survival of such patients.^[12]

Materials and Methods

This study was carried out at the department of Clinical Virology of a tertiary care teaching hospital in South India between 2006 and 2007. Whole blood samples were collected from individuals with HIV-1 established infection who are not receiving antiretroviral therapy (ART). Sixty eight HIV-1 infected individuals who reported to the virology department for the CD4+ T cell estimation and or viral load tests were included in this study. We also collected whole blood samples from 30 normal healthy controls. At the time of sample draw, the HIV-1 infected individuals were not specifically diagnosed to be ill with any opportunistic viral infection by the referring clinician. All the study volunteers were enrolled in the study after a written informed consent for HIV related testing by a

UNAIDS approved HIV antibody test.

Sample collection

Two ml whole blood samples were collected in sterile 5 ml vacutainer (Greiner bio-one, Austria) tubes with K3EDTA as anticoagulant and 8 ml of whole blood were collected in another sterile tube containing 150 µL 10% EDTA from HIV-1 infected individuals by venupuncture between 8.00 to 10.00 AM. The samples were collected in the morning to avoid diurnal variation for the CD4+ T cell estimation. The 400µL of whole blood and separated plasma samples were stored at -80°C in multiple aliquots until testing. A sample of 200 microliters was extracted using QIA amp DNA blood mini reagents (Qiagen, Hilden, Germany) from whole blood samples stored at -80°C. About 200 microliters of eluted DNA was stored at -30°C until testing.

Multiplex PCR

Multiplex PCR was carried out to detect possible opportunistic viruses in HIV-1 infected individuals. Multiplex PCR was standardized using three cocktails or PCR mixes. The detection profile was cocktail-1: CMV, HSV-2 and HSV-1; cocktail-2: VZV, EBV and adenovirus and cocktail-3: JC and BK. The sequences of the primers used including the ones used for in-house HIV-1 and EBV real time PCR are shown in Table 1 and are from published literature.^[13-17] Adenovirus primer amplifies the Hexon

Table 1: Sequences of primers used in the study for multiplex PCR and quantitation real time PCR

Primers	Name	Region	Sequence (5'-3')	Reference
HSV-1	H1P32- FOR	RL 2	TGGGACACATGCCTTCTTGG	13
	H1M32- REV		ACCCCTAGTCAGACTCTGTACTTACCC	
HSV-2	H2M40- FOR	UL 28	GTACAGACCTTCGGAGG	13
	H2P4- REV		CGCTTCATCATGGGC	
CMV	CP15- FOR	IRL 11	GTACACGCACGCTGGTTACC	13
	CM3- REV		GTAGAAAGCCTCGACATCGC	
Adenovirus	Hex3- FOR	Hexon	GACATGACTTTCGAGGTCGATCCCATGGA	14
	Hex4- REV		CCGGCTGAGAAGGGTGTGCGCAGGTA	
EBV	EP5- FOR	Exon 4/5 terminal protein RNA	AACATTGGCAGCAGGTAAGC	13
	EM3- REV		ACTTACCAAGTGTCCATAGGAGC	
VZV	VP22- FOR	ORF 8	CACACGATAATGCCTGATCGG	13
	VM20- REV		TGCTGATATTTCCACGGTACAGC	
Polyomavirus	PV-BACK	VP 1	GGTGCCAACCTATGGAACAG	15
	PV-SNFOR		TCTTTAGTRGTATACACAGCAAA	
JC	JC-FOR	VP 1	GAGGAATGCATGCAGATCTA	15
BK	BK-FOR		ACAGCAAAGCAGGCAAG	
HIV-1	gag- FOR	gag	CATGTTTTTCAGCATTATCAGAAGGA	16
	gag-REV		TGCTTGATGTCCCCCACT	
EBV (in-house Quantitation Real time PCR)	FOR	BNT p143 region	GGAACCTGGTCATCCTTTGC	17
	REV		ACGTGCATGGACCGGTTAAT	

region of 18 serotypes.

Cell culture supernatants were used as the sources of positive controls for viruses like CMV, HSV-1, HSV-2, EBV and adenoviruses. Live attenuated vaccines (Oka strain) were used to extract VZV DNA, while plasmids were used for JC and BK (John Hopkins University, Baltimore, USA). Sterile doubled distilled water was used in PCR testing after every third sample as negative control to check for cross contamination.

For the detection of these eight viruses, their multiplex PCR reactions protocol were as follows: Cocktail-1 (CMV, HSV-2 and HSV-1) was performed using 2.5 units of hot start Taq polymerase (Qiagen), 0.2mM dNTPs and 20 picomoles of primers each in a reaction volume of 50 μ l. Cocktail-2 (VZV, EBV and Adenovirus) was performed using 2.5 units of hot start Taq polymerase (Qiagen), 0.4mM dNTPs and 20 picomoles of primers each in a reaction volume of 50 μ l. Cocktail-3 (JC and BK) was performed using 3.5 units of hot start Taq polymerase (Qiagen), 0.4 mM dNTPs and 25 picomoles of primers each in a reaction volume of 50 μ L. Five microliters of extracted DNA was added to the total reaction volume of 50 μ L for each cocktail.

The multiplex PCR thermal cycling condition for all three cocktails was performed with the initial Taq activation temperature of 95°C for 15 minutes. Subsequently, Cocktail –1 was cycled at 94°C for 30 sec, 52°C for 40 sec, 72°C for 50 sec for 40 cycles and final extension at 78°C for 15 min. Cocktail –2 was cycled at 94°C for 45 sec, 57.5°C for 50 sec, 72°C for 70 sec for 40 cycles and final extension at 72°C for 7 min. Cocktail –3 was a hemi nested PCR with two rounds of amplification. In the first round, amplification cycling conditions were 94°C for 3 min, 94°C for 20 sec, 53°C for 20 sec and 72°C for five min for 30 cycles. The cycling conditions of second round hemi-nested PCR was also the same except for a change in the annealing temperature of 58°C for 20 sec. All the assays for the three cocktails were amplified using thermal cycler Gene amp PCR system 2400 (Perkin Elmer, Massachusetts, USA) or PTC-100 (MJ Research, California, USA).

The amplified products were detected by 2% agarose gel in 1X TBE containing 0.5 μ g/ml ethidium bromide (Sigma, St. Louis, USA) in agarose gel electrophoresis unit. The gels were viewed in a UV Transilluminator (Mighty Bright, Hoefer Scientific Instruments, San Francisco, USA) and photographed using Gel documentation 2000 (Bio Rad, California, USA). The expected specific product sizes for all target genes (amplicons) of the viruses were as follows: Cocktail-1, HSV-1 –147 bp, HSV-2 –227 bp, and CMV 256 bp; Cocktail-2 VZV –275 bp, EBV –182 bp and adenovirus –139 bp and Cocktail-3 JC –199 bp and BK –246 bp respectively. All the samples negative for all the viruses were also tested for α tubulin gene to check the integrity of DNA in the sample. The lower detection limit for the PCR

was estimated for all the eight viruses.

CD4+ T cell estimation

The CD4+ T cell counts for all these HIV-1 infected patients were performed using the standard procedures by (Guava® EasyCD4™ System with CytoSoft™ software, version 2.2 (Guava technologies, California, USA).

In-house HIV-1 RNA PCR

The in-house HIV-1 RNA real time PCR assay was designed to amplify a target gag region and was standardized using the National Institute for Biological Standards and Controls, UK (NIBSC) plasmid standards. QIA amp minelute virus spin assay (Qiagen, GmbH, Germany) was used to extract HIV-1 RNA from plasma samples (in-house HIV-1 RNA assay). Ten microliters of HIV-1 RNA extracts were amplified using real time PCR (Rotor gene RG-3000, Corbett Research, Sydney Australia). The cycling conditions for this in-house HIV-1 RNA assay were 50°C for 30 min and 95°C for 15 min followed by 94°C for 45 sec and 60°C for 75 sec for 45 cycles. The WHO first international standard for HIV-1 RNA 97/656 was used to assess the sensitivity of this in-house HIV-1 RNA real time PCR assay and it was found to be 1000 copies/ml. The standards used in the different assay runs were from a panel standardized against the NIBSC standards or pretested plasma sample using a commercial (Artus Qiagen, GMBH, Germany) assay.

EBV Quantitation PCR

The in-house EBV quantitation real time PCR assay was designed to amplify the BNT p143 region. The standards were prepared using plasmids that were cloned using Topo TA Clone/TOP10 kit. The plasmids were quantitated using the DNA spectrophotometry (BioTek Instruments, Vermont, USA) and then diluted serially in tenfold dilutions of milliQ water to obtain 10⁶ to 10² copies/ μ l. The plasmids were aliquoted and stored at -30°C. Probes for the detection of EBV DNA were labeled with reporter dyes Cy5 and black hole quencher (BHQ) at the 5' and 3' end, respectively (Metabion, Martinsried, Germany, Operon Biotechnologies GmbH, Cologne, Germany). Amplification of EBV DNA was done using 10 μ l of DNA extracts was added to 25 μ l mix containing 12.5 μ l of QuantiTect Multiplex PCR NoROX master mix (Hamburg, Germany) and 0.075 μ l (7.5 picomoles) of EBV primers, 0.05 μ l (5 picomoles) of EBV probe. The thermal cycling conditions used to amplify EBV DNA were as follows 95°C for 15 min, 95°C for 45 sec, and 60°C for 75 sec for 50 cycles. The EBV DNA was amplified and detected using real time PCR (Rotor gene RG-3000, Corbett Research, Sydney Australia).

CMV Quantitation PCR assays

The CMV quantitation was carried out by a

commercially available assay from Qiagen (artus CMV RG PCR assay Hamburg, Germany). This commercial assay used primers targeted 105 bp region of CMV genome. The standards for the CMV PCR were already pre-extracted and supplied by the kit manufacturer. The CMV PCR amplification and detection was done using real time PCR (Rotor Gene RG-3000, Corbett Research, Sydney Australia) as per manufacturer's instructions.

Validation of the PCR assays

The validation of multiplex PCR was carried out by establishing the lower limit of detection and specificity of primers by testing with heterologous viruses. The lower limit of detection was established using titrated virus or plasmids. The PCR amplification was obtained in a virus specific manner. The lower detection limits were 6.25 genome copies/5 μ L for CMV, 0.56 TCID₅₀/5 μ L for HSV-1, 316.2 TCID₅₀/5 μ L for HSV-2, 10 pfu/5 μ L for VZV, 4.3 $\times 10^6$ plasmids/5 μ L for EBV, 0.315 TCID₅₀/5 μ L adenovirus, 1.751 $\times 10^7$ plasmids/5 μ L for JC and 1.8 $\times 10^8$ plasmids/5 μ L for BK. The JC and BK virus PCRs were hemi nested PCRs and the detection limit was estimated using a cloned plasmid preparation which was derived from the PCR product.

Statistical analysis

Chi-square test was used for comparing the percentage positives of EBV and CMV in patients with the two CD4+ T cell groups (less than or equal to 200 and greater than 200 cells/ μ L) and two HIV-1 RNA groups (greater than 100000 and less than 100000).

Results

The 30 normal healthy controls were negative for HIV antibody. Among the 68 samples from HIV-1 infected individuals, 49 (72%) samples were found to be positive for EBV. Nineteen samples were negative by all three cocktails. These 49 samples were repeated again by EBV uniplex PCR to confirm EBV positive status. In 49 EBV positive individuals, 27 had CD4+ T cell counts less than or equal to 200 cells / μ L (range 6-186 cells / μ L) and 22 had CD4+ T cell counts greater than 200 cells / μ L (range of 202-845 cells / μ L). Samples of 5 (7%) individuals were positive for CMV. Among these, three samples showed co-infection with EBV. All the CMV positive patients had a CD4+ T cell count of less than or equal to 200 cells / μ L. The data on EBV and CMV positivies based on the CD4+ T cell counts are shown in Table 2. Among the 30 healthy controls one individual was positive for EBV. The frequency of EBV and CMV positivies in individuals with CD4+ T cell count less than or equal to 200 and greater than 200 cells/ μ L was compared. There was no significant difference between the two groups for EBV positivity ($p = 0.33$). However, for CMV positivity it just fell short of significance ($p = 0.07$). The EBV quantitation PCR results

Table 2: The EBV and CMV positives in individuals stratified based on CD4+ T cell counts.

CD4+ T cell counts (Cells/ μ L)	HIV-1 POS (n=68)	EBV (POS)	CMV (POS)
> 200	33	22	-
≤ 200	35	27	5*

*3 showed co-infection with EBV.

in both HIV-1 infected individuals and control group based on the CD4+ T cell counts are shown in Table 3. All the 49 EBV samples were found to be positive and their EBV DNA load ranged from 300 to 543300 copies/ml. The one control sample were found to have EBV DNA load of 5800 copies/ml. There was no significant difference in the viral load level between the two groups of patients stratified by CD4+ T cell counts into less than or equal to 200 or greater than 200 cells/ μ L. A higher percentage of individuals (77%) who had EBV were found to be having a higher HIV-1 viral load (> 100000 copies/ml). This difference was also not significant ($p = 0.26$). The mean EBV viral load was 3.68 log₁₀ and 3.91log₁₀ among individuals with HIV-1 viral load of less than 100000 RNA copies/ml group and > 100000 copies/ml respectively. This difference was not statistically significant ($p =$ greater than 0.05). The CMV DNA load among the five positive individuals ranged between 80-48141049 copies/ml. In five of the 68 individuals HIV-1, RNA was undetectable. Two among these five individuals had a CD4+ T cell count of greater than 200 cells while the other three had less than or equal to 200 cells. The clinical and laboratory findings of the five CMV positive individuals in the study are given in Table 4.

Discussion

Opportunistic viral infections play a vital role in the progression of the disease in HIV infected individuals. The co-infections with these viruses would require specific treatment when immunosuppression advances. Hence, it is important to detect these opportunistic viruses in HIV infected individuals to reduce morbidity and mortality. Since all HIV infected individuals have to be screened for these viruses, it will be economical to use a multiplex PCR. Our data showed that majority (72%) of the HIV infected individuals were positive for EBV. The percentage positives was found to be higher among individuals with CD4 + T cells less than or equal to 200 cells/ μ L. However, the difference was not significant. In a similar way a higher number of individuals who had EBV showed a HIV-1 viral load of greater than 100,000 copes/ μ L (77% compared to 65%). Subsequent quantitation of the EBV DNA in all the samples failed to find any significant association between EBV load, CD4+ T cells and HIV-1 viral load level. The patients in our study group failed to show evidence of infection with HSV-1, HSV-2, VZV, adenovirus, JC and BK.

Table 3: The EBV load in HIV infected and healthy individuals

Subjects	CD4+ T cell counts	n	EBV (viral load copies/ml) Range	Mean (log10) ± SD
HIV-1 infected	> 200	33	400-543300	3.88±0.64
	≤200	35	300-47500	3.75 ± 0.66
Healthy controls	ND	30	5800	3.76

ND- Not done

Table: 4 Clinical and laboratory findings in individuals with CMV and /or EBV co-infections.

S.no	Study No	CD4+ Tcell count cells/μL	HIV-1 Viral load copies/mL	CMV viral load copies/mL	EBV viral load copies/mL	Cardinal findings
1.	CD 06/323	60	172745	80	34400	Low grade fever, weight loss, Molluscum contagiosum, extrapulmonary tuberculosis, diarrhea, ↑ ALT
2.	CD 06/373	109	65111	1045	Neg	Oral candidiasis, fever.
3.	CD 06/401	15	74445	48141049	15600	Mild hepatomegaly, Molluscum contagiosum, oral candidiasis, ↑ ALT
4.	CD 06/639	71	238778	287	18700	Lymphadenopathy, disseminated tuberculosis, fever, ↑ ALT
5.	CD 07/561	23	795815	5420	Neg	Oesophageal candidiasis, weight loss, cervical tuberculous lymphadenitis, Retinitis

↑ Increased level of liver enzyme

One study looked at the prevalence of opportunistic DNA viruses in saliva of HIV infected individuals and found samples positive for HSV-1, CMV, EBV and HHV-8^[18]. The EBV and HHV-8 were found more frequently even in presence of ART.^[18] They also showed that EBV and CMV were found to be detected significantly more when the CD4+ T cell counts were less than or equal to 200 cells/μL. Another study had reported that EBV and CMV were commonly detected in CSF samples of HIV infected individuals and some co-infection of CMV with EBV was documented.^[19]

In our study group among the five CMV positive individuals, one individual had retinitis while another three had increased ALT level suggesting CMV induced hepatitis. It is important to start pre-emptive anti-CMV therapy in patients who are CMV DNA positive to prevent major CMV associated complications like retinitis. However, one of the arguments in this case is that the CMV is detected in blood and hence it may indicate only a latent infection. It will be ideal to standardize a real time PCR to detect immediate early mRNA (IE mRNA) to find active CMV infection. The detection of IE mRNA for CMV is found to be a useful parameter for initiation of preemptive therapy in transplant

recipients.^[20] Also, this is a laboratory based cross sectional study and hence the in-use utility of the assays could be established only in long term cohort of HIV infected individuals to assess the likelihood ratios of the assays.

Stevens *et al.* (2001) have reported un-fractionated whole blood for EBV detection as the preferred clinical sample in HIV infected individuals, patients with infectious mononucleosis, Burkitt's lymphoma and lung transplant patients.^[11] It is also shown that compared to plasma sample the amount of EBV DNA in whole blood is higher.^[21] The level of EBV DNA in whole blood sample of EBV related symptomatic transplant patients are significantly higher than asymptomatic transplant individuals.^[21] However this difference was not observed when plasma was used as sample.^[21] In our study, we also found high EBV positives when whole blood was tested. The CMV positives in our study were 7.4% and all these individuals had a CD4+ T cell count of less than or equal to 200 cells/μL. These findings are important in light of the earlier report that the presence of CMV and EBV significantly delay the response to ART in HIV-1 infected individuals.^[22]

This study uses the benefits of multiplex PCR which

can amplify many targets in a single reaction tube and with variety of clinical samples. The multiplex PCR has many advantages like saving time, labor, cost effectiveness and ease of performance. One of the drawbacks of our study is the low sensitivity of these assays as the format is essentially a non nested PCR. It is important to increase detection limit of these assays which can be done by doing a nested PCR or a real time PCR with specific probes.^[23] However, the multiplexing is comparatively easy with real time PCR. Another advantage of doing real time PCR is the ability to quantify. It is shown for several viruses like CMV, HHV-8 and EBV that there exists an association between copy number of viruses and clinical illness.^[21,24] Our study showed significant association between the HIV disease stage by CD4+ T cells and CMV positive status but not EBV. Though there is no association of EBV load and HIV disease stage in our population the effect of EBV positives on response to ART has to be assessed.

One study had compared quantitative real time PCR and conventional gel based quantitation PCR assay and obtained very similar results.^[25] In their study they also used whole blood samples but from pediatrics transplant individuals. Real time PCR added advantage over conventional PCR by eliminating both post PCR steps and need for diluting samples like EBV, CMV with high viral titers.^[25] Development of quantitative real time PCR is very useful for diagnosing the disease and also prediction of the development of disease. This will also help to monitor the antiviral treatment response. Screening of HIV patients for other opportunistic viruses may also help in identifying the patients who may develop IRIS with these.^[26]

This is the first report from India on viral opportunistic infections detectable in the blood of HIV infected individuals and demonstrates the high association of EBV and CMV with HIV infection. Some viruses like JC, BK were not detected in whole blood. Previous studies have had success with other appropriate samples like CSF and urine respectively.^[7] More studies are required to fully understand the implications of these findings.

Acknowledgment

Bulk of the study was carried out by Intramural (Christian Medical College, Vellore) support. Quantitation PCR carried out for EBV and CMV was supported from a DBT (Government of India) grant received on September 4, 2008.

References

1. Xu LP, Zhang HY, Huang XJ, Liu KY, Liu DH, Han W, *et al.* Hemorrhagic cystitis following hematopoietic stem cell transplantation: Incidence, risk factors and association with CMV reactivation and graft-versus-host disease. *Chin Med J* 2007;120:1666-71.
2. Musacchio JG, Carvalho Mda G, Morais JC, Silva NH, Scheliga A, Romano S, *et al.* Detection of free circulating Epstein-Barr virus DNA in plasma of patients with Hodgkin's disease. *Sao Paulo Med J* 2006;124:154-7.
3. Hogg RS, Yip B, Chan KJ, Wood E, Craib KJP, O'Shaughnessy MV, *et al.* Rates of disease progression by baseline CD4 cell count and viral load after initiating triple-drug therapy. *JAMA* 2001;286:2568-77.
4. De La Blanchardiere A, Rozenberg F, Caumes E, Picard O, Lionnet F, Livartowski J, *et al.* Neurological complications of varicella-zoster virus infection in adults with human immunodeficiency virus infection. *Scand J Infect Dis* 2000;32:263-9.
5. Schneider U, Ruhnke M, Delecluse HJ, Stein H, Huhn D. Regression of Epstein-Barr virus-associated lymphoproliferative disorders in patients with acquired immunodeficiency syndrome during therapy with foscarnet. *Ann Hematol* 2000;79:214-6.
6. Kagu MB, Nggada HA, Garandawa HI, Askira BH, Durosini MA. AIDS-associated Kaposi's sarcoma in Northeastern Nigeria. *Singapore Med J* 2006;47:1069-74.
7. Elfaitouri A, Hammarin AL, Blomberg J. Quantitative real-time PCR assay for detection of human polyomavirus infection. *J Virol Methods* 2006;135:207-13.
8. Hierholzer JC. Adenoviruses in the immunocompromised host. *Clin Microbiol Rev* 1992; 5:262-74.
9. Marshall DS, Linfert DR, Draghi A., McCarter YS, Tsongalis GJ. Identification of Herpes simplex virus genital infection: Comparison of a Multiplex PCR assay and traditional viral isolation techniques. *Mod Pathol* 2001;14:152-6.
10. Elnifro EM, Ashshi AM, Cooper RJ, Klapper PE. Multiplex PCR: Optimization and application in diagnostic virology. *Clin Microbiol Rev* 2000;13:559-70.
11. Stevens SJ, Pronk I, Middeldrop JM. Toward Standardization of Epstein-barr Virus DNA load monitoring: Unfractionated whole blood as preferred clinical specimen. *J Clin Microbiol* 2001;39:1211-6.
12. Spector SA, Mckinley GF, Lalezari JP, Samo T, Andruczk R, Follansbee S, *et al.* Oral ganciclovir for the prevention of cytomegalovirus disease in persons with AIDS. *N Engl J Med* 1996;334:1491-7.
13. Markoulatos P, Georgopoulou A, Siafakas N, Plakokefalos E, Tzanakaki G, Kourea-Kremastinou J. Laboratory diagnosis of common Herpesvirus infections of the central nervous system by a Multiplex PCR assay. *J Clin Microbiol* 2001;39:4426-32.
14. Echavarria M, Forman M, Ticehurst J, Dumler JS, Charache P. PCR method for detection of Adenovirus in urine of healthy and human immunodeficiency virus-infected individuals. *J Clin Microbiol* 1998;36:3323-6.
15. Biel SS, Held TK, Landt O, Niedrig M, Gelderblom HR, Siegert W, *et al.* Rapid quantification and differentiation of human Polyomavirus DNA in undiluted urine from patients after bone marrow Transplantation. *J Clin Microbiol* 2000;38:3689-95.
16. Palmer S, Wiegand AP, Maldarelli F, Bazmi H, Mican JM, Polis M, *et al.* New real-time reverse transcriptase-initiated PCR assay with single-copy sensitivity for human immunodeficiency virus type -1 RNA in plasma. *J Clin Microbiol* 2003;41:4531-6.
17. Watzinger F, Suda M, Preuner S, Baumgartinger R, Ebner K, Baskova L, *et al.* Real time quantitative PCR assays for detection and monitoring of pathogenic human viruses in immunosuppressed pediatric patients. *J Clin Microbiol* 2004;42:5189-98.
18. Miller CS, Berger JR, Mootoor Y, Avdiushko SA, Zhu H,

- Kryscio RJ. High prevalence of multiple human Herpesviruses in saliva from human immunodeficiency virus-infected persons in the era of highly active Antiretroviral Therapy. *J Clin Microbiol* 2006;44:2409-15.
19. Rojanawiwat A, Miura T, Thaisri H, Pathipvanich P, Umnajsirisuk S, Koibuchi T, *et al.* Frequent detection of Epstein-Barr virus and Cytomegalovirus but not JC virus DNA in cerebrospinal fluid samples from human immunodeficiency virus infected patients in northern Thailand. *J Clin Microbiol* 2005;43:3484-6.
 20. Gerna G, Baldanti F, Lilleri D, Parea M, Alessandrino E, Pagani A, *et al.* Human Cytomegalovirus Immediate-Early mRNA Detection by Nucleic Acid Sequence-Based Amplification as a New Parameter for Preemptive Therapy in Bone Marrow Transplant Recipients. *J Clin Microbiol* 2000;38:1845-53.
 21. Wada K, Kubota N, Ito Y, Yagasaki H, Kato K, Yoshikawa T, *et al.* Simultaneous quantitation of Epstein barr virus, cytomegalovirus and human herpesvirus 6 DNA in samples from transplant recipients by multiplex real time PCR assay. *J Clin Microbiol* 2007;45:1426-32.
 22. Panagiotakis SH, Soufla G, Baritaki S, Sourvinos G, Passam A, Zagoreos I, *et al.* Concurrent CMV and EBV DNAemia is Significantly correlated with a delay in the response to HAART in treatment-naïve HIV type 1-positive patients. *AIDS Res and Hum Retrovir* 2007;23:10-8.
 23. Edwards J, Constable F, Wiechel T, Salib S. Comparison of the molecular tests – single PCR, nested PCR and quantitative PCR (SYBR®Green and TaqMan®) - for detection of *Phaeomoniella chlamydospora* during grapevine nursery propagation. *Phytopathol Mediterr* 2007;46:58-72.
 24. Brantsaeter AB, Holberg-Petersen M, Jeansson S, Goplen AK, Bruun JN. CMV quantitative PCR in the diagnosis of CMV disease in patients with HIV-infection – a retrospective autopsy based study. *BMC Infect Dis* 2007;7:127-34.
 25. Jebbink J, Bai X, Rogers BB, Dawson DB, Scheuermann RH, Domiati-Saad R. Development of real time PCR assays for the quantitative detection of Epstein-Barr virus and cytomegalovirus, comparison of TaqMan probes, and molecular beacons. *J Mol Diagn* 2003;5:15-20.
 26. Murdoch DM, Venter WDF, Rie AV, Feldman C. Immune reconstitution inflammatory syndrome (IRIS): review of common infectious manifestations and treatment options. *AIDS Res Ther* 2007;4:9.

Source of Support: Study was carried out by Intramural (Christian Medical College, Vellore) support. Quantitation PCR carried out for EBV and CMV was supported from a DBT (Government of India) grant received on September 4, 2008.

Conflict of Interest: None declared.

